

Microbiome report

Sample ID 33333333
Sampling 24/03/2025
Analysed 04/04/2025

Material Stool
Laboratory labors.at

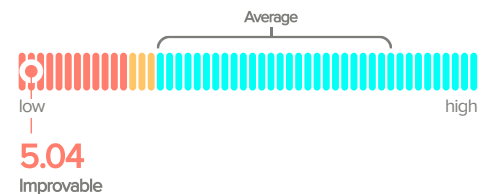
1. Microbial health

Microbial diversity

The overall diversity of your gut bacteria could be better. Our tips will help you strengthen your gut microbiome and enhance your health.

Species richness: 203 (Average: 202-322)

Species evenness: 0.66 (Average: 0.72-0.78)



Additional information

The microbial diversity describes the variety of your gut microbiome and is made up of species richness and species evenness. Diversity is the most important parameter for analysing the health of your gut microbiome. It measures how many different bacterial species (species richness) are present in the gut and how evenly they are distributed among the individual species (species evenness).

A bacterial community with high diversity therefore consists of many different species and the individuals are evenly distributed.

Diversity is calculated using the "Shannon-Index" - the number you see in the graphic. This index takes into account both species richness and evenness (the distribution of individuals among those species). The higher the value, the better!

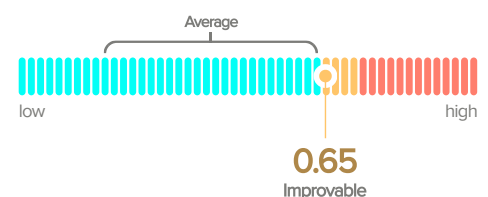
Research shows that low diversity can be linked to various health problems such as inflammatory bowel disease, obesity, metabolic disorders and autoimmune diseases, among others.

Species richness: Indicates the number of different bacterial species in a stool sample. A high value indicates a high species richness.

Species evenness: Provides information on how balanced the frequency of the different types of bacteria is in your gut. A more even distribution helps to prevent the dominance of a few species, making the microbial community less susceptible to disturbances or adverse changes.

Dysbiosis index

Your gut microbiome is out of balance, indicating dysbiosis. Our tips can help you address this issue, strengthen your gut bacteria, and restore balance to your gut microbiome.



Additional information

Dysbiosis refers to an imbalance in your gut microbiome, where potentially harmful bacteria outnumber beneficial ones. This imbalance can result from various factors, including an unbalanced diet high in ultra-processed foods, lifestyle habits such as chronic stress, lack of physical activity, or insufficient sleep, and the regular use of medication. Certain medical conditions, particularly inflammatory bowel disease, can also increase the risk of dysbiosis. The dysbiosis index quantifies the severity of this imbalance and can be useful for monitoring changes in the microbiome during treatments or dietary adjustments.

Enterotype

Enterotype 3 ("Ruminococcus") is associated with a diet that is balanced and rich in complex carbohydrates, including fibre.



Enterotyp 3: Ruminococcus

Additional information

Enterotype categorise the gut microbiome into three dominant bacterial groups, which form the "basic microbiome" during the early years of life, primarily influenced by genetic factors and dietary habits. There is evidence that your enterotype may influence which foods you can metabolise particularly well and how efficient vitamin production occurs in your gut. Long-term dietary habits, along with age, health status, and medication use, can influence your enterotype.

Please note that this classification and your result indicate only a tendency and the types may overlap.

Enterotype 3

Enterotype 3 is characterised by a predominance of the bacterial genus Ruminococcus. This enterotype is particularly common among individuals with a mixed diet. Ruminococcus bacteria quickly and effectively break down indigestible food components and then convert them into health-promoting substances such as short-chain fatty acids. Ruminococcus bacteria produce more enzymes that can break down indigestible carbohydrates such as fibre and then convert them into energy. However, these bacteria can also harm the intestinal mucosa by breaking down sugar-protein complexes present in it.

Enterotype 1

Enterotype 1 ("Bacteroides") is associated with a diet rich in animal foods.

Enterotype 2

Enterotype 2 ("Prevotella") is associated with a plant-based diet rich in fruits, vegetables, legumes and whole grains.

2. Gut-body interaction

Gut-immune axis

High level of support for your immune system



The composition of your gut bacteria seems to support your immune system well.

Additional information

The gut-immune axis describes the connection and interaction between the gut and the body's immune system. Over 70% of the immune system is located in the gut and is supported by the bacteria living there. Certain bacteria activate immune cells or regulate their activity and produce anti-inflammatory substances such as short-chain fatty acids. The health of your gut microbiome is crucial for a strong immune system.

A stool sample can provide valuable insights into the strength and resilience of your immune system.

Inflammatory potential

There might be inflammation present in your gut. Your inflammation potential is increased. Our tips can help you to increase the number and activity of anti-inflammatory bacteria.



High inflammatory potential

Your result is made up of 4 metabolic pathways for LPS production:

-  Sugar building blocks for LPS
-  Extended LPS modules
-  Surface antigens
-  Preliminary stage for LPS

Additional information

The inflammatory potential in your gut can be calculated using **lipopolysaccharides (LPS)**. LPS are molecules that occur in the cell walls of certain bacteria. Recent research shows how the composition of the gut microbiome (including the types and amounts of LPS-producing bacteria) influences the immune system in the gut.

Certain bacteria can use the metabolic pathways described to produce LPS. If there are too many LPS-producing bacteria in your gut, the potential for inflammation is increased.

In addition, the dysbiosis index and immune strength are important parameters that can influence the inflammatory potential in the gut.

Gut-skin axis

Your gut microbiome seems to support healthy skin.

Low tendency to skin conditions



Low tendency to acne



Low tendency to neurodermatitis



High tendency to psoriasis



Additional information

The gut-skin axis describes the connection between the gut microbiome and skin health. Skin diseases such as acne, neurodermatitis and psoriasis are often caused by inflammation in the body, which then becomes visible on the surface of the skin. Intestinal bacteria can directly regulate the immune system and inflammatory processes in the body. It is therefore possible to draw conclusions about skin health by analysing the bacteria found in a stool sample.

Weight management

Your intestinal bacteria could help you regulate your weight. If you are pursuing health goals, seek additional individual advice from a healthcare professional.

Low tendency to underweight



High tendency to overweight



Additional information

The composition of gut bacteria influences various aspects of metabolism, such as energy production from food. Several studies indicate that gut bacteria play a role in weight regulation. Some bacteria are associated with a lean physique, while others may contribute to obesity.

Your gut may harbour both types of bacteria associated with being underweight and those linked to being overweight.

Other important factors in weight regulation include microbial diversity and short-chain fatty acids produced by bacteria.

3. Gut health

Leaky gut syndrome

High tendency to leaky gut syndrome



The composition of your gut bacteria indicates a possible leaky gut syndrome. If you suffer from digestive problems or symptoms, we recommend that you consult a doctor.

Additional information

Leaky gut syndrome describes an increased permeability of the intestinal mucosa. It controls which substances pass from the gut into the bloodstream. If the intestinal mucosa is impaired, unwanted substances can enter the body and trigger inflammation. To maintain an intact intestinal mucosa, a healthy gut microbiome is key. Beneficial gut bacteria help strengthen the gut barrier and reduce inflammation. Please note that a high diversity is crucial for maintaining a healthy intestinal mucosa.

On the other hand, if your gut bacteria are imbalanced, certain bacteria can proliferate that break down mucosal cells excessively, making the intestinal mucosa more "leaky". This can lead to leaky gut syndrome. An impaired intestinal mucosa is associated with an increased risk of chronic inflammation, food intolerances, autoimmune diseases, irritable bowel syndrome and skin diseases, among other things.

Irritable bowel syndrome

Low tendency to irritable bowel syndrome



There seems to be no connection between your gut bacteria and irritable bowel syndrome.

Additional information

Irritable bowel syndrome (IBS) is a common gastrointestinal disorder characterised by symptoms such as diarrhoea and/or constipation, bloating, and abdominal pain. Studies show that affected individuals often have an unfavourable composition and lower diversity of gut bacteria compared to individuals without IBS. There are numerous causes that can trigger irritable bowel syndrome or exacerbate symptoms. Psychological factors, such as stress, appear to play a particularly important role. In addition, malnutrition, nutrient deficiencies, other diseases, toxins, a lack of stomach acid, medication, infections and an imbalance in the gut microbiome are among the potential triggers.

SIBO

High tendency to SIBO



Your gut microbiome indicates a possible bacterial overgrowth in the small intestine. If you are experiencing symptoms, we recommend consulting a doctor.

Additional information

SIBO (Small Intestinal Bacterial Overgrowth) refers to an excessive growth of bacteria in the small intestine. Typically, the small intestine contains far fewer bacteria than the large intestine. In cases of SIBO, however, there is an increased presence of bacteria from the large intestine in the small intestine. This is usually triggered by slower digestion or anatomical changes following surgery. The most common symptoms are bloating, but other digestive disorders and nutrient deficiencies (especially vitamin B12) can also occur.

Research findings show that bacterial overgrowth in the small intestine also has an impact on the composition of the bacteria in the large intestine. A stool sample can therefore provide indications of the possible presence of SIBO. If the result is positive, consider consulting a doctor and carrying out an additional breath test.

Gluten sensitivity

Low tendency to gluten sensitivity



The composition of your gut microbiome provides no evidence of gluten sensitivity. Foods containing gluten do not appear to cause you any symptoms.

 Additional information

Gluten sensitivity refers to a reaction to gluten that is not associated with celiac disease (an autoimmune response) or a wheat allergy. Gluten is a protein found in grains such as wheat, spelt, rye and barley. The consumption can lead to digestive issues and symptoms such as chronic fatigue and headaches in individuals with gluten sensitivity.

Based on scientific studies, it is now possible to establish a link between the composition of the gut microbiome and the potential for gluten sensitivity.

4. Well-being

Mental well-being

High tendency to mental imbalance



Your results indicate an imbalance in the gut-brain axis, which may affect mood, stress response, or cognitive function. Our tips can help nurture your microbiome and support better mental well-being.

 Additional information

There is a strong connection between mental health and the intestinal microbiome, often referred to as the "gut-brain axis". This means that the bacteria in your gut can communicate with your brain and influence its function. Certain gut bacteria can produce substances like neurotransmitters, metabolites, and hormones that send signals to the brain and influence its function. Changes in their levels can affect mood and play a role in regulating social behavior, depressive-like behavior, and motivation.

The communication between the gut and the brain is bidirectional. This means that not only does the gut influence the brain, but the brain can also impact the gut. For instance, stress or strong emotional states can lead to changes in gut function and disrupt the balance of the gut bacteria.

Physical well-being

High tendency to physical imbalance



Your gut bacteria are showing signs of imbalance, which could be affecting your physical fitness and vitality. Our tips can help you to rebalance your microbiome and support your physical well-being.

 Additional information

Regular physical activity can positively influence the composition of your gut microbiome by increasing the diversity of gut bacteria and supporting the growth of health-promoting bacteria that produce short-chain fatty acids. The relationship between exercise and the gut microbiome is reciprocal: not only does physical activity influence the microbiome, but the composition of the gut flora also contributes to exercise performance. Studies have shown that changes in the microbiome through exercise can help to prevent metabolic diseases, strengthen the immune system and promote general well-being. An active lifestyle is therefore crucial to support gut health and physical wellbeing.

5. Metabolic health

Blood sugar regulation

High tendency to unstable blood sugar regulation



Your gut microbiome could better support your insulin sensitivity and blood sugar regulation. Our tips can help strengthen your gut bacteria and improve your blood sugar balance.

Additional information

The gut microbiome plays a crucial role in regulating blood sugar levels by influencing metabolism, insulin sensitivity, and inflammation. Certain beneficial bacteria help break down dietary fibres into short-chain fatty acids (SCFAs), which improve insulin sensitivity and reduce blood sugar spikes. Additionally, the gut microbiome communicates with the endocrine system, influencing the secretion of hormones such as glucagon-like peptide-1 (GLP-1), which enhances insulin release and helps maintain stable blood sugar levels. An imbalance in gut bacteria (dysbiosis) is associated to an increased risk of insulin resistance, type 2 diabetes, and metabolic disorders. By nurturing a diverse and healthy gut microbiome, you can positively impact blood sugar regulation and overall well-being.

Blood pressure regulation

High tendency to dysregulated blood pressure regulation



Your gut microbiome could better support your blood pressure regulation. Our tips can help rebalance your gut bacteria and support heart health.

Additional information

A healthy gut microbiome plays a crucial role in blood pressure regulation. Studies suggest that hypertension may be associated with an imbalanced gut flora and a weakened intestinal barrier, allowing harmful substances to enter the body and trigger inflammation, which further raises blood pressure. Certain bacteria that produce short-chain fatty acids (SCFAs) and have anti-inflammatory effects are often less abundant in people with hypertension. Additionally, gut microbes influence key factors involved in blood pressure regulation, such as angiotensin and nitric oxide signaling.

Gut-liver axis

High tendency to an imbalance in the gut-liver axis



The composition of your gut bacteria indicates a potential imbalance in your gut-liver axis. By following our tips, you can support a healthy microbiome and gut barrier, which in turn helps promote optimal liver function.

Additional information

The gut-liver axis refers to the close connection between the microbiome in the gut and the health of the liver. The two organs are directly connected via the portal vein, which transports substances from the gut to the liver. This direct route means that an imbalance in the gut can have a significant impact on the liver - and vice versa.

Liver diseases such as fatty liver and inflammation are often linked to gut dysbiosis - where there is often a reduction in microbial diversity, an increase in potentially harmful bacteria and a decrease in beneficial bacteria that produce short-chain fatty acids (SCFAs). When the intestinal barrier is weakened, more harmful substances can enter the bloodstream and reach the liver. This can overload the protective functions of the liver and lead to inflammation, an increased immune response, liver damage and an increased risk of infection. This in turn can further increase the permeability of the intestinal barrier and have a negative impact on the microbiome - creating a vicious circle.

Thyroid activity

Your gut bacteria appear to support healthy thyroid function by contributing to nutrient absorption, hormone conversion, and immune balance.



Additional information

The gut microbiome is closely linked to thyroid activity, particularly through its influence on the immune system. Gut bacteria play a central role in regulating the immune system, which is instrumental in the development of autoimmune thyroid diseases - such as Hashimoto's thyroiditis (often associated with hypothyroidism) and Graves' disease (often associated with hyperthyroidism).

Changes in the microbiome can lead to a disruption in immune regulation and inflammation, thereby promoting hypothyroidism. It can also impair the absorption of important nutrients such as iodine, selenium, iron and zinc, which are essential for the production of thyroid hormones.

Our own data show that hyperthyroidism in particular appears to result in a loss of microbial diversity in the gut, which significantly impairs immune function and the stability of the intestinal barrier. Reduced diversity could unbalance the immune system to such an extent that it leads to thyroid overactivity.

6. Bacteria lists

F/B ratio



Bacterium	Frequency (%)	Reference (%)
Bacteroidota	41.57	30.88 - 45.11
Firmicutes	52.46	48.28 - 63.47

Probiotic bacteria

Bacterium	Frequency (%)	Reference (%)
Akkermansia muciniphila	0.00	0.00 - 1.56
Bifidobacterium	0.45	0.03 - 0.66
Bifidobacterium longum	0.19	0.00 - 0.36
Lactobacillus	0.00	0.00 - 0.01

Mucin-producing bacteria

Bacterium	Frequency (%)	Reference (%)
Akkermansia muciniphila	0.00	0.00 - 1.56
Bacteroides fragilis	0.22	0.00 - 0.26

Bacterium	Frequency (%)	Reference (%)
Bacteroides thetaiotaomicron	↑ 0.92 	0.02 - 0.65
Bifidobacterium	0.45 	0.03 - 0.66
Faecalibacterium prausnitzii	9.56 	3.44 - 11.31
Lactobacillus	0.00 	0.00 - 0.01

Butyrate-producing bacteria

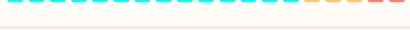
Bacterium	Frequency (%)	Reference (%)
Anaerostipes	↑ 0.45 	0.04 - 0.36
Coprococcus	↓ 0.00 	0.06 - 1.90
Eubacterium hallii group	↑ 0.36 	0.00 - 0.09
Faecalibacterium prausnitzii	9.56 	3.44 - 11.31
Roseburia	↑ 0.36 	0.00 - 0.16
Subdoligranulum	1.90 	0.30 - 2.50

Sulphate-reducing bacteria

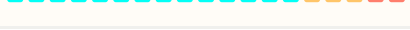
Bacterium	Frequency (%)	Reference (%)
Bilophila	0.17 	0.02 - 0.25
Bilophila wadsworthia	0.17 	0.00 - 0.24
Desulfovibrio	0.00 	0.00 - 0.23

Overview of all bacteria

Phylum	Genus	Frequency (%)	Reference (%)
Firmicutes	Acetanaerobacterium	0.01 	No data
Firmicutes	Agathobacter	0.34 	No data
Verrucomicrobiota	Akkermansia	0.00 	0.00 - 1.94
Bacteroidota	Alistipes	2.48 	1.35 - 4.96
Firmicutes	Anaerofilum	0.00 	0.00 - 0.01
Firmicutes	Anaerostipes	↑ 0.45 	0.04 - 0.36
Firmicutes	Anaerotruncus	0.00 	0.00 - 0.01
Bacteroidota	Bacteroides	↑ 36.65 	11.89 - 31.62
Bacteroidota	Barnesiella	↓ 0.00 	0.13 - 2.04

Phylum	Genus	Frequency (%)	Reference (%)
Actinobacteriota	Bifidobacterium	0.45 	0.03 - 0.66
Desulfobacterota	Bilophila	0.17 	0.02 - 0.25
Firmicutes	Blautia	0.00 	0.00 - 0.16
Firmicutes	Butyricicoccus	0.11 	0.11 - 0.47
Bacteroidota	Butyricimonas	↑ 0.72 	0.00 - 0.22
Firmicutes	CAG-56	0.00 	0.00 - 0.12
Firmicutes	Christensenellaceae R-7 group	↓ 0.00 	0.05 - 1.83
Firmicutes	Clostridium sensu stricto 1	0.00 	0.00 - 0.11
Firmicutes	Colidextribacter	↑ 0.34 	0.05 - 0.19
Actinobacteriota	Collinsella	↓ 0.00 	0.02 - 0.22
Bacteroidota	Coprobacter	0.00 	0.00 - 0.24
Firmicutes	Coprococcus	↓ 0.00 	0.06 - 1.90
Firmicutes	Defluviitaleaceae UCG-011	↑ 0.04 	0.00 - 0.03
Desulfobacterota	Desulfovibrio	0.00 	0.00 - 0.23
Firmicutes	Dialister	0.00 	0.00 - 2.50
Firmicutes	Dielma	↑ 0.05 	0.00 - 0.01
Firmicutes	DTU089	↑ 0.03 	0.00 - 0.02
Actinobacteriota	Eggerthella	0.03 	No data
Firmicutes	Eisenbergiella	↑ 0.56 	0.00 - 0.02
Actinobacteriota	Enterorhabdus	↑ 0.02 	0.00 - 0.01
Firmicutes	Erysipelatoclostridium	↑ 0.06 	0.00 - 0.05
Firmicutes	Erysipelotrichaceae UCG-003	↑ 0.53 	0.03 - 0.33
Proteobacteria	Escherichia-Shigella	0.00 	0.00 - 0.07
Firmicutes	Eubacterium eligens group	↓ 0.00 	0.25 - 2.68
Firmicutes	Eubacterium hallii group	↑ 0.36 	0.00 - 0.09
Firmicutes	Eubacterium nodatum group	0.00 	0.00 - 0.02
Firmicutes	Eubacterium oxidoreducens group	0.00 	0.00 - 0.05
Firmicutes	Eubacterium ruminantium group	0.00 	0.00 - 0.13
Firmicutes	Eubacterium siraeum group	↑ 1.86 	0.01 - 1.49
Firmicutes	Eubacterium ventriosum group	0.03 	0.01 - 0.15

Phylum	Genus	Frequency (%)	Reference (%)
Firmicutes	Eubacterium xylanophilum group	0.00 	0.00 - 0.37
Firmicutes	Faecalibacterium	↑ 27.32 	6.02 - 15.78
Firmicutes	Family XIII AD3011 group	↑ 0.12 	0.00 - 0.08
Firmicutes	Family XIII UCG-001	↑ 0.07 	0.00 - 0.03
Firmicutes	Flavonifractor	↑ 0.28 	0.00 - 0.06
Firmicutes	Fournierella	0.00 	0.00 - 0.01
Firmicutes	GCA-900066575	0.02 	0.01 - 0.08
Proteobacteria	Haemophilus	0.00 	0.00 - 0.12
Firmicutes	Holdemanella	0.00 	0.00 - 0.09
Firmicutes	Holdemania	↑ 0.07 	0.00 - 0.05
Firmicutes	Hungatella	0.03 	No data
Firmicutes	Hydrogenoanaerobacterium	0.00 	0.00 - 0.01
Firmicutes	Intestinibacter	0.02 	0.00 - 0.03
Firmicutes	Intestinimonas	↑ 0.18 	0.00 - 0.06
Firmicutes	Lachnoclostridium	↑ 1.78 	0.15 - 0.82
Firmicutes	Lachnospira	0.91 	0.35 - 2.63
Firmicutes	Lachnospiraceae FCS020 group	↓ 0.00 	0.03 - 0.37
Firmicutes	Lachnospiraceae NC2004 group	↓ 0.00 	0.02 - 1.00
Firmicutes	Lachnospiraceae ND3007 group	↓ 0.00 	0.24 - 2.58
Firmicutes	Lachnospiraceae NK4A136 group	0.00 	0.00 - 0.28
Firmicutes	Lachnospiraceae UCG-001	↓ 0.00 	0.02 - 0.51
Firmicutes	Lachnospiraceae UCG-004	0.05 	0.00 - 0.29
Firmicutes	Lachnospiraceae UCG-008	0.00 	0.00 - 0.05
Firmicutes	Lachnospiraceae UCG-010	↓ 0.00 	0.03 - 0.27
Firmicutes	Lactobacillus	0.00 	0.00 - 0.01
Firmicutes	Lactococcus	0.00 	0.00 - 0.01
Firmicutes	Marvinbryantia	0.00 	0.00 - 0.02
Firmicutes	Merdibacter	0.00 	0.00 - 0.01
Firmicutes	Monoglobus	↓ 0.00 	0.07 - 0.37
Firmicutes	Moryella	0.00 	0.00 - 0.05

Phylum	Genus	Frequency (%)	Reference (%)
Firmicutes	Negativibacillus	0.05 	0.00 - 0.08
Firmicutes	NK4A214 group	0.00 	0.00 - 0.95
Bacteroidota	Odoribacter	↑ 0.64 	0.11 - 0.41
Firmicutes	Oscillibacter	↑ 0.61 	0.03 - 0.31
Firmicutes	Oscillospira	0.08 	0.00 - 0.11
Proteobacteria	Oxalobacter	0.00 	0.00 - 0.03
Firmicutes	Paludicola	2.16 	No data
Bacteroidota	Parabacteroides	1.09 	0.94 - 3.59
Bacteroidota	Paraprevotella	0.00 	0.00 - 0.36
Proteobacteria	Parasutterella	0.17 	0.01 - 0.80
Firmicutes	Peptococcus	0.00 	0.00 - 0.03
Firmicutes	Phascolarctobacterium	1.23 	0.00 - 2.07
Firmicutes	Phoceia	↑ 0.01 	0.00 - 0.01
Bacteroidota	Prevotella	0.00 	0.00 - 11.55
Firmicutes	Pseudoflavonifractor	↑ 0.11 	0.00 - 0.01
Firmicutes	Romboutsia	0.00 	0.00 - 0.01
Firmicutes	Roseburia	↑ 0.36 	0.00 - 0.16
Firmicutes	Ruminococcus	↓ 0.00 	0.15 - 1.95
Firmicutes	Ruminococcus gauvreauii group	0.03 	No data
Firmicutes	Ruminococcus torques group	↑ 0.16 	0.00 - 0.11
Firmicutes	Sellimonas	0.01 	No data
Actinobacteriota	Senegalimassilia	0.00 	0.00 - 0.02
Actinobacteriota	Slackia	0.00 	0.00 - 0.02
Firmicutes	Streptococcus	0.04 	0.03 - 0.31
Firmicutes	Subdoligranulum	1.90 	0.30 - 2.50
Proteobacteria	Sutterella	↑ 4.04 	0.02 - 2.45
Firmicutes	Terrisporobacter	0.00 	0.00 - 0.02
Patescibacteria	TM7x	0.01 	0.00 - 0.03
Firmicutes	Turicibacter	0.00 	0.00 - 0.06
Firmicutes	Tuzzerella	0.02 	No data

Phylum	Genus	Frequency (%)	Reference (%)
Firmicutes	Tyzzarella	0.00 	0.00 - 0.11
Firmicutes	UBA1819	↑ 0.11 	0.00 - 0.05
Firmicutes	UCG-002	↓ 0.00 	0.03 - 0.63
Firmicutes	UCG-003	0.09 	0.00 - 0.18
Firmicutes	UCG-005	↓ 0.00 	0.02 - 0.54
Firmicutes	UCG-009	0.00 	0.00 - 0.01
Firmicutes	Veillonella	0.00 	0.00 - 0.15
Verrucomicrobiota	Victivallis	0.00 	0.00 - 0.24

7. Recommendations

Microbial health

Include fermented foods

Fermented foods provide probiotics that support a balanced gut microbiome, reduce dysbiosis, and enhance digestion. They diversify gut flora, strengthen the gut lining, and lower gut permeability. Regular consumption also boosts immunity and combats inflammation, promoting overall health.

Integrate fermented foods into your meals at least 3-4 times a week:

- (plant based) Yogurt and kefir: Start your day with these in smoothies or in your muesli. Enjoy them as a snack with berries and nuts.
- Sauerkraut and kimchi: Add these tangy side dishes to your meals for extra flavor and nutrition. Opt for vegan kimchi if you're allergic to fish.
- Kombucha: Swap sugary drinks for kombucha as a refreshing alternative.
- Miso: Add miso to soups or use it as a flavor enhancer in dressings. Miso comes in varieties made from soybeans, chickpeas, barley, or rice.

Enterotype 3 – ruminococcus dominant

Enterotype 3 is associated with a mixed diet that includes both animal and plant-based foods. Ruminococcus species are efficient at breaking down resistant starch, so incorporating these starches is beneficial for gut health. A well-rounded diet combining plant-based and animal proteins is key to supporting this enterotype.

Strengthen your microbiome by incorporating resistant starch and varied protein sources:

- Resistant starch: Include cooled potatoes, rice, and legumes in your meals regularly. Resistant starch is also found in cashews, raw oats, and slightly ripe bananas.
- Combine plant and animal proteins: Mix and match proteins for optimal protein utilization. For example: Egg and beans with spinach and potatoes, Beans with sweetcorn, Lentil dal with brown rice, Hummus with wholemeal bread.
- Healthy fats and hydration: Consume beneficial fats like olive oil, sesame seeds, and avocado. Drink plenty of water to prevent constipation, a common issue for this enterotype.

Increase fiber intake

Fiber-rich foods support a diverse microbiome by nourishing beneficial bacteria. Dietary fiber acts as a prebiotic, enhances microbial diversity, reduces dysbiosis, and promotes digestive health. A variety of fiber sources improves bowel regularity and helps prevent gut issues.

Consume a variety of fiber-rich foods daily:

- **Whole grains:** Incorporate oats, quinoa, and brown rice into your meals.
- **Legumes:** Add beans, lentils, and chickpeas to salads, soups, and stir-fries.
- **Fruits:** Enjoy apples, pears, and berries as snacks or desserts.
- **Vegetables:** Include broccoli, carrots, and spinach in your main dishes or as side options.

Berry millet breakfast bowl

Ingredients:

1 cup cooked millet
1/2 cup mixed berries (blueberries, strawberries, blackberries)
1 tablespoon chia seeds
1 cup almond milk
1-2 teaspoons honey or nut butter (optional)

Preparation:

1. Place cooked millet in a bowl.
2. Add mixed berries and chia seeds on top.
3. Pour almond milk over the mixture.
4. Drizzle with honey or nut butter if desired.

A delightful start to your day, packed with fiber and antioxidants.

Gut-body interaction

Choose healthy fats

Healthy fats, particularly Omega-3 fatty acids, are vital for reducing inflammation and supporting a balanced gut microbiome. These fats contribute to maintaining the integrity of the gut lining and are associated with better skin hydration and texture. Including a variety of healthy fat sources in your diet can enhance nutrient absorption and support the growth of beneficial gut bacteria.

Include healthy fats in your meals regularly:

- **Fatty fish:** Make salmon, mackerel, or sardines a regular part of your diet.
- **Nuts:** Snack on a handful of almonds, pistachios or walnuts, or add them to oatmeal or yogurt for extra crunch.
- **Seeds:** Sprinkle chia seeds or flaxseeds into smoothies or yogurt for added nutrition and fibre.
- **Avocado:** Incorporate avocado into salads or sandwiches, or enjoy as a spread on whole-grain bread.

Antibiotic awareness

Antibiotics can promote the growth of pathogenic bacteria, such as *Fusobacterium*, which can cause inflammation in the body. Unfortunately, antibiotics do not only kill harmful bacteria but also beneficial species.

Discuss antibiotic use with your doctor carefully:

- **Evaluate Necessity** with your doctor: Consider whether antibiotics are truly necessary for your condition.
- **Discuss Alternatives** with your doctor: Explore alternative treatments that may be effective without antibiotics.
- **Post-Antibiotic Care:** Consider probiotics or fermented foods after antibiotic treatment to restore gut health.

Boost b vitamins

Ensure adequate intake of B vitamins to support gut health and promote anti-inflammatory benefits.

Incorporate these B vitamin-rich foods:

- **Sunflower Seeds:** A great source of B vitamins, perfect for snacking or adding to salads.
- **Eggs:** A versatile ingredient that can be included in various meals for a protein boost and B vitamins.
- **Spinach and Leafy Greens:** Rich in essential nutrients, these can be used in salads, smoothies, or cooked dishes.
- **Chickpeas:** A fantastic legume that provides B vitamins and can be used in salads, hummus, or soups.

Quinoa veggie stir fry

Ingredients:

1 cup quinoa
2 cups vegetable broth
1 cup broccoli florets
1 cup chopped bell peppers
1 cup sliced carrots
1 tablespoon olive oil
2 cloves garlic, minced
1 tablespoon soy sauce
1 teaspoon ginger, grated

Preparation:

1. Rinse quinoa under cold water.
2. Bring vegetable broth to a boil and add quinoa. Reduce heat, cover, and simmer for 15 minutes.
3. In a pan, heat olive oil and sauté garlic and ginger until fragrant.
4. Add broccoli, bell peppers, and carrots to the pan, stir-frying for 5-7 minutes until tender.
5. Mix in cooked quinoa and soy sauce, stirring well to combine all ingredients.
6. Serve warm, garnished with fresh herbs if desired.

A nourishing stir fry with a perfect blend of protein-rich quinoa and colorful veggies.

Intestinal health

Focus on micronutrients

Ensuring sufficient intake of vitamins, antioxidants, and polyphenols is crucial for maintaining the integrity of the gut barrier. A deficiency in these nutrients may lower the abundance of *A. muciniphila*, which is essential for gut health.

Focus on nutrient-rich foods:

- **Berries:** Blueberries, raspberries, and acai berries are packed with antioxidants and vitamins—perfect for snacks or breakfast additions.
- **Leafy Greens:** Incorporate nutrient-dense greens like kale and spinach into your meals for added health benefits.
- **Healthy Nuts and Seeds:** Walnuts, flaxseeds, and almonds provide essential nutrients and healthy fats—add them to your salads or smoothies.

Manage stress levels

Chronic stress can significantly impact your gut health and exacerbate symptoms like digestive discomfort, bloating, and imbalances. By incorporating stress-reduction techniques, you can foster a more harmonious gut environment. Addressing stress through mindfulness and promoting calm can have positive effects on your microbiome, supporting a better balance of gut bacteria.

Practice stress-reduction techniques regularly:

- **Yoga:** Participate in regular yoga classes to improve relaxation.
- **Meditation:** Set aside time for meditation or deep-breathing exercises.
- **Mindful Eating:** Pay attention to your meals, chewing slowly, and enjoying the flavors.
- **Nature Walks:** Spend time outdoors to relax and unwind.
- **Hobbies:** Engage in activities you enjoy to alleviate stress.

Incorporating pseudograins

Pseudograins are naturally gluten-free and rich in fiber, magnesium, iron, and essential vitamins. They offer a nutritious alternative to traditional grains and are often better tolerated by people with sensitive digestion.

Include nutrient-dense pseudograins in your meals to support digestive health:

- Add quinoa as a side dish or in salads and curries.
- Use amaranth for breakfast porridge, in soups, or puffed in muesli.
- Enjoy chia seeds in puddings, smoothies, or even in water for added fiber.
- Incorporate buckwheat flour into pancakes or homemade bread for a gluten-free option.

Miso and vegetable stir-fry

Ingredients:

2 tablespoons miso paste
3 cups mixed vegetables (carrots, broccoli, bell pepper)
1 tablespoon olive oil
2 spring onions (green part only), sliced
1 tablespoon soy sauce
2 tablespoons water

Preparation:

1. In a small bowl, mix miso paste, soy sauce, and water.
2. Heat olive oil in a pan, add spring onions, and sauté till fragrant.
3. Add vegetables and cook until tender-crisp.
4. Pour in miso mixture, stirring to coat the vegetables.
5. Cook for an additional minute and serve hot.

A savory dish brimming with gut-friendly probiotics.

Well-being

Keep to regular bedtimes

A regular sleep rhythm is crucial for optimal serotonin production and general well-being. As the gut microbiome and the brain are closely connected, a regular night's sleep can help to maintain a balance in the gut and improve communication between the gut and brain. Good and sufficient sleep promotes the body's regeneration processes and reduces stress hormones, which can have a negative effect on the intestinal environment. Regular bedtimes therefore play an important role in supporting the gut-brain axis.

Some points that should be considered are:

- Avoid caffeine and heavy meals before going to bed.
- Create a relaxing sleeping environment without disturbances.
- Stick to regular bedtimes, even at weekends.
- Avoid using electronic devices just before going to bed.
- Introduce calming rituals such as reading or meditation before going to bed.

Try intermittent fasting

Intermittent fasting - i.e. deliberately planned breaks between meals - can have a positive effect on gut health and the microbiome in particular. These targeted breaks give the beneficial intestinal bacteria time to regenerate, which can strengthen their diversity and function. During this resting phase, the bacteria can recover and perform their tasks - such as supporting digestion, immune defense and communication with the brain - even more effectively. If intermittent fasting is implemented carefully and individually adapted, it can promote balance in the gut and contribute to greater vitality, inner balance and well-being in the long term.

Take in enough vitamin c

Vitamin C plays a crucial role in supporting the production of neurotransmitters, which are essential for communication between neurons in the brain. Ensuring adequate levels of vitamin C can significantly improve the ability to concentrate. As a powerful antioxidant, vitamin C also protects nerve cells from damage caused by free radicals. It also promotes the absorption of iron, which can optimize the brain's oxygen supply.

Note the following tips:

- Start with a 16:8 cycle, i.e. fast for 16 hours and eat for 8 hours.
- Avoid processed foods during the eating phase.
- Drink plenty of water to support fasting in the best possible way.
- Pay attention to your body's signals and adjust your cycle if necessary.

Points to note:

- Increase your consumption of fruit and vegetables rich in vitamin C, such as citrus fruits, peppers and broccoli.
- Do not overcook food or eat it raw to minimize the loss of vitamin C.
- Ensure sufficient sleep and good stress management to maintain the vitamin C concentration in the body.

Potato salad with resistant starch and herbs

Ingredients:

500g cooked and cooled potatoes
2 tablespoons olive oil
1 tablespoon apple cider vinegar
Salt and pepper to taste
Fresh herbs: parsley, dill and chives

Preparation:

1. Cut the potatoes into even slices.
2. Whisk together the olive oil, apple cider vinegar, salt and pepper in a large bowl.
3. Add the potatoes to the mixture and stir gently until they are well coated.
4. Garnish with fresh herbs and serve.

The use of cooked and chilled potatoes increases their resistant starch content and promotes intestinal health and general well-being by supporting healthy bacteria.

Metabolic health

Consume foods rich in polyphenols

Polyphenols - natural plant substances - make a valuable contribution to the health of the gut-liver axis. They specifically promote the growth of helpful intestinal bacteria such as *Bacteroides eggerthii* and thus support a balanced, diverse microbiome. Thanks to their strong antioxidant and anti-inflammatory properties, polyphenols not only have a stabilizing effect on the intestinal flora, but also relieve the liver in its daily detoxification processes. A diet rich in polyphenol-containing foods strengthens microbial diversity and balances the finely tuned interaction between the gut and liver.

Note the following points:

- Include blueberries and raspberries in your daily breakfast, e.g. in porridge or (plant-based) yogurt. They are rich in polyphenols and support liver health.
- Drink a cup of green tea every day. Green tea is rich in polyphenols and has an anti-inflammatory effect, which can support the liver.
- Enjoy 1-2 pieces of dark chocolate. Choose varieties with at least 85% cocoa, as these have a higher polyphenol content.

Avoid highly processed foods

Highly processed foods and sugar promote the growth of pro-inflammatory bacteria such as *Bacteroides vulgatus* and *Bacteroides stercoris*. An imbalance in the microbiome can exacerbate the symptoms of hyperthyroidism and impair thyroid function. It is therefore crucial to reduce the consumption of such foods. Instead, the diet should be fresh and balanced, with a focus on fruits, vegetables and whole grains to support the microbiome and promote anti-inflammatory bacteria.

Pay attention to the following points:

- Avoid ready meals and sweets, as they often contain high amounts of additives, sugar, sweeteners and refined carbohydrates.
- Go for fresh, unprocessed foods. Instead of a classic breakfast like sugary cereal or toast, try baked sweet potatoes with avocado and lime or avocado toast with egg and optionally some chili. For a probiotic version, you can enjoy (plant-based) yogurt with nuts, seeds and cinnamon.
- Try to cook for yourself as often as possible and plan your meals in advance so that you have healthy food to hand during the working day.

Minimize alcohol consumption

Alcohol can affect the balance of the gut-liver axis by weakening the intestinal barrier and disrupting the microbial balance in the gut. This can promote the colonization of unfavorable bacteria, which in turn can put a strain on the liver. Reducing your alcohol intake - ideally to one or two drinks a week or no alcohol at all - helps to keep the microbiome in a healthy state. This promotes the growth of beneficial bacteria that not only support gut health, but also relieve the liver in a natural way.

Pay attention to the following points:

- Choose non-alcoholic alternatives. There is now a wide range of non-alcoholic wines and beers that offer the taste of alcohol without the side effects.
- Try Kombucha: A fermented drink with a slight alcohol content, but without the typical negative effects of alcohol. It has a refreshing taste and also aids digestion.
- Enjoy sparkling water with lemon, lime, ginger, mint leaves or some fruit juice: a simple but delicious option. The tangy feeling and fruity taste can easily replace the "alcohol moment".

Blueberry and pomegranate yogurt

Ingredients:

200 ml natural yogurt or a plant-based alternative
100 g fresh blueberries
½ fresh pomegranate, seeded
1 tsp honey
50 g walnuts, chopped

Preparation:

1. Put the yogurt in a bowl.
2. Spread the blueberries and pomegranate seeds over the yogurt.
3. Drizzle the honey over the top.
4. Sprinkle with the chopped walnuts.

This blueberry and pomegranate yogurt is rich in polyphenols, which promote the growth of beneficial bacteria and can therefore protect your liver.

Summary

Here's a short breakdown of how to boost your gut health:

- **Embrace fermented foods:** Fermented foods provide beneficial probiotics that support a balanced gut microbiome, reduce dysbiosis, and enhance digestion. Regularly consuming them also boosts immunity and combats inflammation, promoting overall health by strengthening the gut lining and diversifying gut flora.
- **Diversify your diet:** A mixed diet that includes both animal and plant-based foods, especially resistant starches, is beneficial for gut health. This approach supports a balanced enterotype 3, enhancing microbial diversity and digestive function.
- **Increase fiber intake:** Fiber-rich foods nourish beneficial bacteria, acting as a prebiotic to enhance microbial diversity, reduce dysbiosis, and promote digestive health. A varied intake of fiber improves bowel regularity and helps prevent gut issues.
- **Prioritize vitamins, antioxidants, and polyphenols:** Sufficient intake of vitamins, antioxidants, and polyphenols is vital for maintaining the gut barrier and supporting beneficial bacteria like *A. muciniphila*. These nutrients also protect nerve cells and optimize brain oxygen supply.

- **Manage chronic stress:** Chronic stress negatively impacts gut health, exacerbating symptoms like discomfort and bloating. Implementing stress-reduction techniques, such as mindfulness, can promote a harmonious gut environment and better balance of gut bacteria.
- **Opt for pseudograins:** Pseudograins are naturally gluten-free and rich in fiber, magnesium, iron, and essential vitamins. They serve as a nutritious alternative to traditional grains, often better tolerated by those with sensitive digestion.
- **Include healthy fats:** Healthy fats, especially omega-3s, are crucial for reducing inflammation and supporting a balanced gut microbiome. They maintain gut lining integrity, enhance nutrient absorption, and promote beneficial gut bacteria.
- **Limit antibiotics:** Antibiotics can disrupt the gut microbiome by eliminating both harmful and beneficial bacteria, potentially promoting the growth of pathogenic species that cause inflammation. Discuss antibiotic use with your doctor.
- **Ensure sufficient b vitamin intake:** Adequate intake of B vitamins is crucial for supporting gut health and promoting anti-inflammatory benefits, contributing to overall gut and systemic well-being.
- **Maintain a regular sleep schedule:** A consistent sleep rhythm is vital for optimal serotonin production and overall well-being. It supports a balanced gut microbiome and enhances the gut-brain axis communication, promoting bodily regeneration and reducing stress hormones.
- **Consider intermittent fasting:** Intermittent fasting, with deliberate breaks between meals, can positively impact gut health by allowing beneficial bacteria to regenerate, enhancing their diversity and function. This promotes gut balance and overall vitality.
- **Increase vitamin c intake:** Vitamin C supports neurotransmitter production, improving concentration, and acts as an antioxidant, protecting nerve cells. It also boosts iron absorption, optimizing brain oxygen supply.
- **Integrate polyphenols into your diet:** Polyphenols contribute to gut-liver axis health by promoting beneficial bacteria and aiding liver detoxification. Their antioxidant and anti-inflammatory properties stabilize gut flora, fostering a balanced microbiome.
- **Avoid highly processed foods and sugar:** These foods promote pro-inflammatory bacteria and can exacerbate issues like hyperthyroidism and impair thyroid function. Focus on fresh, balanced meals with fruits, vegetables, and whole grains to support your microbiome.
- **Reduce alcohol consumption:** Alcohol can weaken the gut barrier and disrupt microbial balance, potentially promoting unfavorable bacteria and straining the liver. Aim for minimal or no alcohol to support a healthy microbiome and liver function.

8. About us

At Superhuman, we believe that peak vitality, performance and a long, healthy life shouldn't be left to chance—they should be your reality. As Serbia's first dedicated longevity centre, we are here to help you unlock your body's full potential, ensuring you don't just live longer, but feel younger and more energized every single day. Our approach is built on strictly science-based protocols and cutting-edge diagnostics, moving beyond generic health trends to deliver results that are highly personalized to your unique biology. By bridging the gap between world-class research and your daily well-being, we empower you to take definitive, data-driven control of your health. Add years to your life and life to your years!

9. Questionnaire

Personal

Is this test and questionnaire for someone else?

No

When is your birthday?

1982-10-10

What is your biological sex?

Male

How tall are you?

188 cm

How much do you weigh?

99 kg

In which country were you born?

AT

In which country do you currently reside?

AT

Which region of the world better describes your ancestry?

Eastern Europe

Were you born by C-section?

No

Your sample

When did you take your sample?

2025-03-24

What is the stool consistency of your sample?

4

Nutrition

Do you follow a specific diet?

Yes

Which diet(s) do you follow?

Ovo-Lacto-Vegetarian

When did you last take probiotics?

Over a year ago

How often do you drink alcohol?

Less than 1x per week

Lifestyle

How many people live in your household?

3-5 people

Do you have any pets living with you?

Yes

Which type of pets?

Dog(s)

Do you smoke?

Occasionally

How many hours do you sleep on average per night?

7-9 hours

How would you assess your current stress level?

2

Have you been outside of Europe in the last 6 months?

No

Gut health

How often do you have bowel movements on average?

1-3x per day

Do you have regular discomfort in the gastrointestinal tract?

No

Do you have any allergies?

No

Do you suffer from any intolerances?

No

Do you suffer from a chronic inflammatory bowel disease?

No

Have you been diagnosed with any other digestive tract disease?

No

How long has it been since your last colonoscopy?

Never

General health

How would you rate your physical health?

6

How would you assess your psychological well-being?

4

Do you have skin problems?

No

Do you suffer from any infectious disease(s)?

No

Do you have any mental or neurological disorders?

Yes

Please select the appropriate mental or neurological disorder(s)

Depression

Do you have any of the following diseases?

No

Drugs

When did you last take antibiotics?

Over a year ago

Are you regularly taking any medication?

Yes

Which of the following medications do you take?

Anti-depressants

Please note

Detection of a microorganism by this test does not imply the presence of a disease. Similarly, non-detection of a microorganism by this test does not rule out the presence of a disease-causing microorganism. Other organisms may also be present that are not detected by this test. This test is not a substitute for established methods for the identification of microorganisms or their antimicrobial susceptibility profile.

The analysed data is examined using specific phylogenetic analysis algorithms to obtain precise results on the basis of which your microbiome report is created. The latest scientific findings, bioinformatic excellence and AI-supported algorithms are used for this. These machine learning algorithms are used, with the highest standards of privacy and data security, in the gut health and gut-body interaction (excluding inflammatory potentials) chapters to determine the propensity of the microbiome profile of a sample to the profile of samples from individuals with specific characteristics. The summary in the chapter "Recommendations" was created with the help of AI.

We accept no liability for health decisions made on the basis of the test results.

References

- [1] Human Microbiome Project Consortium. A framework for human gut microbiota. *Nature*. 2012.
- [2] Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. *Nature*. 2012.
- [3] Lozupone CA, et al. Diversity, stability and resilience of the human gut microbiota. *Nature*. 2012.
- [4] Yatsunenko T, et al. Human gut microbiome viewed across age and geography. *Nature*. 2012.
- [5] Langille MG, et al. Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nat Biotechnol*. 2013.
- [6] Ranjan R, et al. Multiomic strategies reveal diversity and important functional aspects of human gut microbiome. *Biomed Res Int*. 2018.
- [7] King CH, et al. Baseline human gut microbiota profile in healthy people and standard reporting template. *PLoS ONE*. 2019.
- [8] Martinez-Guryn M, et al. Regional diversity of the gastrointestinal microbiome. *Cell Host Microbe*. 2019.
- [9] Young VB, et al. Overview of the gastrointestinal microbiota. *Adv Exp Med Biol*. 2008.
- [10] Chatelier E, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013.
- [11] Cotillard A, et al. Dietary intervention impact on gut microbial gene richness. *Nature*. 2013.
- [12] Li J, et al. An integrated catalog of reference genes in the human gut microbiome. *Nat Biotechnol*. 2014.
- [13] Donaldson GP, et al. Gut biogeography of the bacterial microbiota. *Nat Rev Microbiol*. 2015.
- [14] Pasolli E, et al. Extensive unexplored human microbiome diversity revealed by over 150,000 genomes from metagenomes spanning age, geography, and lifestyle. *Cell*. 2019.
- [15] Hagerly SL, et al. An empirically derived method for measuring human gut microbiome alpha diversity: Demonstrated utility in predicting health-related outcomes among a human clinical sample. *PLoS One*. 2020.
- [16] Manor O, et al. Health and disease markers correlate with gut microbiome composition across thousands of people. *Nat Commun*. 2020.
- [17] Kers JG, et al. The power of microbiome studies: Some considerations on which alpha and beta metrics to use and how to report results. *Front Microbiol*. 2022.
- [18] Gevers D, et al. The treatment-naïve microbiome in new-onset Crohn's disease. *Cell Host Microbe*. 2014.
- [19] Casén C, et al. Deviations in human gut microbiota: a novel diagnostic test for determining dysbiosis in patients with IBS or IBD. *Aliment Pharmacol Ther*. 2015.
- [20] Wang J, et al. Gut microbial dysbiosis is associated with altered hepatic functions and serum metabolites in chronic hepatitis B patients. *Front Microbiol*. 2017.
- [21] Farup PG, et al. Separating "good" from "bad" faecal dysbiosis—evidence from two cross-sectional studies. *BMC Obes*. 2018.
- [22] Valeur J, et al. Exploring gut microbiota composition as an indicator of clinical response to dietary FODMAP restriction in patients with irritable bowel syndrome. *Dig Dis Sci*. 2018.
- [23] Wei S, et al. Determining gut microbial dysbiosis: a review of applied indexes for assessment of intestinal microbiota imbalances. *Appl Environ Microbiol*. 2021.
- [24] Malinen E, et al. Association of symptoms with gastrointestinal microbiota in irritable bowel syndrome. *World J Gastroenterol*. 2010.
- [25] Arumugam M, et al. Enterotypes of the human gut microbiome. *Nature*. 2011.
- [26] Wu GD, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science*. 2011.
- [27] Koren O, et al. A guide to enterotypes across the human body: meta-analysis of microbial community structures in human microbiome datasets. *PLoS Comput Biol*. 2013.
- [28] Ley RE, et al. Gut microbiota in 2015: Prevotella in the gut: choose carefully. *Nat Rev Gastroenterol Hepatol*. 2016.
- [29] Mobeen F, et al. Enterotype variations of the healthy human gut microbiome in different geographical regions. *Bioinformation*. 2018.
- [30] Heeney DD, et al. Intestinal Lactobacillus in health and disease, a driver or just along for the ride? *Curr Opin Biotechnol*. 2018.
- [31] Costea PI, et al. Enterotypes in the landscape of gut microbial community composition. *Nat Microbiol*. 2018.
- [32] Naumova N, et al. Human gut microbiome response to short-term Bifidobacterium-based probiotic treatment. *Indian J Microbiol*. 2020.
- [33] Markowiak-Kopeć P, et al. The effect of probiotics on the production of short-chain fatty acids by human intestinal microbiome. *Nutrients*. 2020.
- [34] Bułgryn I, et al. Absence of enterotypes in the human gut microbiomes reanalyzed with non-linear dimensionality reduction methods. *PeerJ*. 2023.
- [35] Zheng D, et al. Interaction between microbiota and immunity in health and disease. *Cell Research*. 2020.
- [36] Robinson I, et al. Assessing the performance of a novel stool-based microbiome test that predicts response to first line immune checkpoint inhibitors in multiple cancer types. *MDPI Cancers*. 2023.
- [37] Di Lorenzo F, et al. Lipopolysaccharide structure of Gram-negative populations in the gut microbiota and effects on host interactions. *FEMS Microbiol Rev*. 2019.
- [38] Kaczmarczyk M, et al. The gut microbiota is associated with the small intestinal paracellular permeability and the development of the immune system in healthy children during the first two years of life. *J Transl Med*. 2021.
- [39] Si J, et al. Gut microbiome signatures distinguish type 2 diabetes mellitus from non-alcoholic fatty liver disease. *Comput Struct Biotechnol J*. 2021.
- [40] Qin Q, et al. The relationship between osteoporosis and intestinal microbes in the Henan Province of China. *Front Cell Dev Biol*. 2021.
- [41] Ghosh SS, et al. Intestinal barrier dysfunction, LPS translocation, and disease development. *J Endocr Soc*. 2022.
- [42] Khorsand B, et al. Overrepresentation of Enterobacteriaceae and Escherichia coli is the major gut microbiome signature in Crohn's disease and ulcerative colitis; a comprehensive metagenomic analysis of IBDMDB datasets. *Front Cell Infect Microbiol*. 2022.
- [43] Nzabarushimana E, et al. Functional profile of host microbiome indicates Clostridioides difficile infection. *Gut Microbes*. 2022.
- [44] Peng YC, et al. Gut microbiome dysbiosis in patients with hepatitis B virus-related hepatocellular carcinoma after extended hepatectomy liver failure. *Ann Transl Med*. 2022.
- [45] Wallen ZD, et al. Metagenomics of Parkinson's disease implicates the gut microbiome in multiple disease mechanisms. *Nat Commun*. 2022.
- [46] Yan H, et al. Gut microbiome alterations in patients with visceral obesity based on quantitative computed tomography. *Front Cell Infect Microbiol*. 2022.
- [47] Zhang WH, et al. Fecal microbiota transplantation ameliorates active ulcerative colitis by downregulating pro-inflammatory cytokines in mucosa and serum. *Front Microbiol*. 2022.
- [48] Di Vincenzo F, et al. Gut microbiota, intestinal permeability, and systemic inflammation: a narrative review. *Intern Emerg Med*. 2023.
- [49] Mena-Vázquez N, et al. Adiposity is associated with expansion of the genus Dialister in rheumatoid arthritis patients. *Biomed Pharmacother*. 2023.
- [50] Patumcharoenpol P, et al. Exploring longitudinal gut microbiome towards metabolic functional changes associated in atopic dermatitis in early childhood. *Biology (Basel)*. 2023.
- [51] Squillario M, et al. Gut-microbiota in children and adolescents with obesity: inferred functional analysis and machine-learning algorithms to classify microorganisms. *Sci Rep*. 2023.
- [52] Tap J, et al. Global branches and local states of the human gut microbiome define associations with environmental and intrinsic factors. *Nat Commun*. 2023.
- [53] Penders J, et al. Molecular fingerprinting of the intestinal microbiota of infants in whom atopic eczema was or was not developing. *Clin Exp Allergy*. 2006.
- [54] Candela M, et al. Unbalance of intestinal microbiota in atopic children. *BMC Microbiol*. 2012.
- [55] Eppinga H, et al. Similar depletion of protective Faecalibacterium prausnitzii in psoriasis and inflammatory bowel disease, but not in hidradenitis suppurativa. *J Crohn's Colitis*. 2016.
- [56] Tang MF, et al. Eczema susceptibility and composition of fecal microbiota at 4 weeks of age: a pilot study in Chinese infants. *Br J Dermatol*. 2016.
- [57] Zheng H, et al. Altered gut microbiota composition associated with eczema in infants. *PLoS One*. 2016.
- [58] Vaughn AR, et al. Skin-gut axis: the relationship between intestinal bacteria and skin health. *World J Dermatol*. 2017.
- [59] Tan L, et al. The Akkermansia muciniphila is a gut microbiota signature in psoriasis. *Exp Dermatol*. 2018.
- [60] Yan HM, et al. Gut microbiota alterations in moderate to severe acne vulgaris patients. *J Dermatol*. 2018.
- [61] Hidalgo-Cantabrana C, et al. Gut microbiota dysbiosis in a cohort of patients with psoriasis. *Br J Dermatol*. 2019.
- [62] Myers B, et al. The gut microbiome in psoriasis and psoriatic arthritis. *Best Pract Res Clin Rheumatol*. 2019.
- [63] Salem I, et al. The gut microbiome as a major regulator of the gut-skin axis. *Front Microbiol*. 2019.
- [64] Visser M, et al. Bacterial dysbiosis and translocation in psoriasis vulgaris. *Front Cell Infect Microbiol*. 2019.
- [65] Chen L, et al. Skin and gut microbiome in psoriasis: gaining insight into the pathophysiology and finding novel therapeutic strategies. *Front Microbiol*. 2020.
- [66] Chen YJ, et al. An altered fecal microbial profiling in rosacea patients compared to matched controls. *J Formos Med Assoc*. 2020.
- [67] Dei-Cas I, et al. Metagenomic analysis of gut microbiota in non-treated plaque psoriasis patients stratified by disease severity: development of a new Psoriasis-Microbiome Index. *Sci Rep*. 2020.
- [68] Hsu D, et al. Role of skin and gut microbiota in the pathogenesis of psoriasis, an inflammatory skin disease. *Microecol Med*. 2020.
- [69] Yegorov S, et al. Psoriasis is associated with elevated IL-1 α and intestinal microbiome alterations. *Front Immunol*. 2020.
- [70] De Pessemer B, et al. Gut-skin axis: current knowledge of the interrelationship between microbial dysbiosis and skin conditions. *Microorganisms*. 2021.
- [71] Olejniczak-Staruch I, et al. Alterations of the skin and gut microbiome in psoriasis and psoriatic arthritis. *Int J Mol Sci*. 2021.
- [72] Park DH, et al. Comparative analysis of the microbiome across the gut-skin axis in atopic dermatitis. *Int J Mol Sci*. 2021.
- [73] Mahmud MR, et al. Impact of gut microbiome on skin health: gut-skin axis observed through the lenses of therapeutics and skin diseases. *Gut Microbes*. 2022.
- [74] Lee MJ, et al. Disordered development of gut microbiome interferes with the establishment of the gut ecosystem during early childhood with atopic dermatitis. *Gut Microbes*. 2022.
- [75] Liu C, et al. Correlation analysis between gut microbiota characteristics and melasma. *Front Microbiol*. 2022.
- [76] Thye AYK, et al. Gut-skin axis: unravelling the connection between the gut microbiome and psoriasis. *Biomedicines*. 2022.
- [77] Ley RE, et al. Microbial ecology: human gut microbes associated with obesity. *Nature*. 2006.
- [78] Turnbaugh PJ, et al. A core gut microbiome in obese and lean twins. *Nature*. 2009.
- [79] Schwertz A, et al. Microbiota and SCFA in lean and overweight healthy subjects. *Obesity*. 2010.
- [80] Clarke SF, et al. The gut microbiota and its relationship to diet and obesity. *Gut Microbes*. 2012.
- [81] Ahmad R, et al. Gut permeability and mucosal inflammation: bad, good or context dependent. *Mucosal Immunol*. 2017.
- [82] Menni C, et al. Gut microbiome diversity and high-fiber intake are related to lower long-term weight gain. *Int. Journal of Obesity*. 2017.
- [83] Del Chierico F, et al. Gut microbiota markers in obese adolescent and adult patients: age-dependent differential patterns. *Front Microbiol*. 2018.
- [84] Gao X, et al. Body mass index differences in the gut microbiota are gender-specific. *Frontiers in Microbiology*. 2018.

- [85] Hjorth MF, et al. Prevotella to Bacteroides ratio predicts body weight and fat loss success on 24-week diets varying in macronutrient composition and dietary fiber: results from a post-hoc analysis. *Int. Journal of Obesity*. 2018.
- [86] Heianza Y, et al. Changes in gut microbiota-related metabolites and long-term successful weight loss in response to weight-loss diets: The POUNDS Lost Trial. *Diabetes Care*. 2018.
- [87] Frost F, et al. A structured weight loss program increases gut microbiota phylogenetic diversity and reduces levels of Collinsella in obese type 2 diabetics: a pilot study. *PLoS ONE*. 2019.
- [88] de Clercq NC, et al. Weight gain after fecal microbiota transplantation in a patient with recurrent underweight following clinical recovery from anorexia nervosa. *Psychother Psychosom*. 2019.
- [89] Camody RN, et al. Roles of the gut microbiome in weight management. *Nature Review Microbiology*. 2023.
- [90] van Hul M, et al. The gut microbiota in obesity and weight management: microbes as friends or foe? *Nat Rev Endocrinol*. 2023.
- [91] Martel J, et al. Gut barrier disruption and chronic disease. *Cell Trends Endocrinol Metab*. 2024.
- [92] Quévrain E, et al. Identification of an anti-inflammatory protein from *Faecalibacterium prausnitzii*, a commensal bacterium deficient in Crohn's disease. *Gut*. 2016.
- [93] Hiippala K, et al. The potential of gut commensals in reinforcing intestinal barrier function and alleviating inflammation. *Nutrients*. 2018.
- [94] Ghosh S, et al. Regulation of intestinal barrier function by microbial metabolites. *Cell Mol Gastroenterol Hepatol*. 2021.
- [95] Kinashi T, et al. Partners in leaky gut syndrome: intestinal dysbiosis and autoimmunity. *Front Immunol*. 2021.
- [96] Swidsinski A, et al. Spatial organization and composition of the mucosal flora in patients with inflammatory bowel disease. *J Clin Microbiol*. 2005.
- [97] Malinen E, et al. Analysis of the fecal microbiota of irritable bowel syndrome patients and healthy controls with real-time PCR. *Am J Gastroenterol*. 2005.
- [98] Kassinen A, et al. The fecal microbiota of irritable bowel syndrome patients differs significantly from that of healthy subjects. *Gastroenterology*. 2007.
- [99] Lyra A, et al. Diarrhea-predominant irritable bowel syndrome distinguishable by 16S rRNA gene phylotype quantification. *World J Gastroenterol*. 2009.
- [100] Rajčić-Stojanović M, et al. Global and deep molecular analysis of microbiota signatures in fecal samples from patients with irritable bowel syndrome. *Gastroenterology*. 2011.
- [101] Jeffery IB, et al. An irritable bowel syndrome subtype defined by species-specific alterations in fecal microbiota. *Gut*. 2012.
- [102] Jalanka-Tuovinen J, et al. Faecal microbiota composition and host-microbe cross-talk following gastroenteritis and in post-infectious irritable bowel syndrome. *BMJ Gut*. 2014.
- [103] Rajčić-Stojanović M, et al. Intestinal microbiota and diet in IBS: causes, consequences, or epiphenomena? *Am J Gastroenterol*. 2015.
- [104] Lopetuso LR, et al. Gut microbiota in health, diverticular disease, irritable bowel syndrome, and inflammatory bowel diseases: time for microbial markers of gastrointestinal disorders. *Dig Dis*. 2017.
- [105] Vich Vila A, et al. Gut microbiota composition and functional changes in inflammatory bowel disease and irritable bowel syndrome. *Sci Transl Med*. 2018.
- [106] Enck P, et al. Dysbiosis in functional bowel disorders. *Ann Nutr Metab*. 2018.
- [107] Rizzello F. Dietary geraniol ameliorates intestinal dysbiosis and relieves symptoms in irritable bowel syndrome patients: a pilot study. *BMC Complement Altern Med*. 2018.
- [108] Hod K, et al. The effect of a multispecies probiotic on microbiota composition in a clinical trial of patients with diarrhea-predominant irritable bowel syndrome. *Neurogastroenterol Motil*. 2018.
- [109] Fodor A, et al. Rifaximin is associated with modest, transient decreases in multiple taxa in the gut microbiota of patients with diarrhea-predominant irritable bowel syndrome. *Gut Microbes*. 2019.
- [110] Pittayanon R, et al. Gut microbiota in patients with irritable bowel syndrome—A systematic review. *Gastroenterology*. 2019.
- [111] Agnello M, et al. Gut microbiome composition and risk factors in a large cross-sectional IBS cohort. *BMJ Open Gastroenterol*. 2020.
- [112] Masoodi I, et al. Microbial dysbiosis in irritable bowel syndrome: a single-center metagenomic study in Saudi Arabia. *JGH Open*. 2020.
- [113] Yang M, et al. Mucosal-associated microbiota other than luminal microbiota has a close relationship with diarrhea-predominant irritable bowel syndrome. 2020.
- [114] Barandouzi Z, et al. Altered gut microbiota in irritable bowel syndrome and its association with food components. *J Pers Med*. 2021.
- [115] Baldelli V, et al. The role of Enterobacteriaceae in gut microbiota dysbiosis in inflammatory bowel diseases. *Microorganisms*. 2021.
- [116] Ghosh S, et al. Regulation of intestinal barrier function by microbial metabolites. *Cell Mol Gastroenterol Hepatol*. 2021.
- [117] Kinashi, et al. Partners in leaky gut syndrome: intestinal dysbiosis and autoimmunity. *Front Immunol*. 2021.
- [118] Sciaivilla P, et al. Gut microbiota profiles and characterization of cultivable fungal isolates in IBS patients. *Applied Microbiology and Biotechnology*. 2021.
- [119] Pimentel M, et al. ACG clinical guideline: Small intestinal bacterial overgrowth. *The American Journal of Gastroenterology*. 2020.
- [120] Ghosal U, et al. Small intestinal bacterial overgrowth in patients with inflammatory bowel disease: A case-control study. *Indian Journal of Gastroenterology*. 2021.
- [121] Banaszak M, et al. Association between gut dysbiosis and the occurrence of SIBO, LIFO, SIFO and IMO. *Microorganisms*. 2023.
- [122] Knez E, et al. The importance of food quality, gut motility and microbiome in SIBO development and treatment. *Nutrition*. 2024.
- [123] Lu S, et al. Differences in clinical manifestations and the fecal microbiome between irritable bowel syndrome and small intestinal bacterial overgrowth. *Digestive and Liver Disease*. 2024.
- [124] Hansen LBS, et al. A low-gluten diet induces changes in the intestinal microbiome of healthy Danish adults. *Nature*. 2018.
- [125] Cardoso-Silva D, et al. Intestinal barrier function in gluten-related disorders. *Nutrients*. 2019.
- [126] Volta U, et al. Nonceliac wheat sensitivity: An immune-mediated condition with systemic manifestations. *Gastroenterology Clinics of North America*. 2019.
- [127] Nobel YR, et al. Lack of effect of gluten challenge on fecal microbiome in patients with celiac disease and non-celiac gluten sensitivity. *Clinical and Translational Gastroenterology*. 2021.
- [128] Clapp M, Aurora N, Herrera L, Bhatia M, Wilen E, Wakefield S. Gut microbiota's effect on mental health: The gut-brain axis. *Clin Pract*. 2017 Sep 15;7(4):987.
- [129] Shoubridge, A.P., Choo, J.M., Martin, A.M. et al. The gut microbiome and mental health: advances in research and emerging priorities. *Mol Psychiatry* 27, 1908–1919 (2022).
- [130] Skonieczna-Zydecka, K.; Marlicz, W.; Misera, A.; Koulaouzidis, A.; Loniewski, I. Microbiome—The Missing Link in the Gut-Brain Axis: Focus on Its Role in Gastrointestinal and Mental Health. *J. Clin. Med.* 2018, 7, 521.
- [131] Moloney, R.D.; Desbonnet, L.; Clarke, G. et al. The microbiome: stress, health and disease. *Mamm Genome* 25, 49–74 (2014).
- [132] Loh, J.S.; Mak, W.Q.; Tan, L.K.S. et al. Microbiota–gut–brain axis and its therapeutic applications in neurodegenerative diseases. *Sig Transduct Target Ther* 9, 37 (2024).
- [133] Verma A, Inslicht SS, Bhargava A. Gut-Brain Axis: Role of Microbiome, Metabolomics, Hormones, and Stress in Mental Health Disorders. *Cells*. 2024 Aug 27;13(17):1436. doi: 10.3390/cells13171436. PMID: 39273008; PMCID: PMC11394554.
- [134] Chen CY, Wang YF, Lei L, Zhang Y. Impacts of microbiota and its metabolites through gut-brain axis on pathophysiology of major depressive disorder. *Life Sci*. 2024 Aug 15;351:122815. doi: 10.1016/j.lfs.2024.122815. Epub 2024 Jun 10. PMID: 38866215.
- [135] Monda V, Villano I, Messina A, Valenzano A, Esposito T, Moscatelli F, Viggiano A, Cibelli G, Chieffi S, Monda M, Messina G. Exercise Modifies the Gut Microbiota with Positive Health Effects. *Oxid Med Cell Longev*. 2017;2017:3831972.
- [136] Cataldi S, Bonavalontà V, Poli L, Clemente FM, De Candia M, Carvutto R, Silva AF, Badicu G, Greco G, Fischetti F. The Relationship between Physical Activity, Physical Exercise, and Human Gut Microbiota in Healthy and Unhealthy Subjects: A Systematic Review. *Biology (Basel)*. 2022 Mar 21;11(3):479.
- [137] Clauss M, Gérard P, Mosca A and Leclerc M (2021) Interplay Between Exercise and Gut Microbiome in the Context of Human Health and Performance. *Front. Nutr.* 8:637010.
- [138] Dzielwiecka, H., Buttar, H.S., Kasperska, A. et al. Physical activity induced alterations of gut microbiota in humans: a systematic review. *BMC Sports Sci Med Rehabil* 14, 122 (2022).
- [139] Fontana, F., Longhi, G., Tarracchini, C. et al. The human gut microbiome of athletes: metagenomic and metabolic insights. *Microbiome* 11, 27 (2023).
- [140] Yun S, Seo Y, Lee Y, Lee DT. Gut microbiome related to metabolic diseases after moderate-to-vigorous intensity exercise. *J Exerc Sci Fit*. 2024 Oct;22(4):375-382. doi: 10.1016/j.jesf.2024.07.003. Epub 2024 Jul 25. PMID: 39185003; PMCID: PMC11342187.
- [141] Ghaffar T, Ubaldi F, Volpini V, Valeriani F, Romano Spica V. The Role of Gut Microbiota in Different Types of Physical Activity and Their Intensity: Systematic Review and Meta-Analysis. *Sports (Basel)*. 2024 Aug 14;12(8):221. doi: 10.3390/sports12080221. PMID: 39195597; PMCID: PMC11360093.
- [142] Fontana F, Longhi G, Tarracchini C, Mancabelli L, Lugli GA, Alessandri G, Turroni F, Milani C, Ventura M. The human gut microbiome of athletes: metagenomic and metabolic insights. *Microbiome*. 2023 Feb 14;11(1):27. doi: 10.1186/s40168-023-01470-9. PMID: 36782241; PMCID: PMC9926762.
- [143] Diet-gut microbiome interaction and its impact on host blood glucose homeostasis: a series of nutritional n-of-1 trials Yu, Y., Yuanqing et al. *eBioMedicine*, Volume 111, 105483.
- [144] Palmäs-Bédard MSA, Costabile G, Vetrani C, Åberg S, Hjalmarsson Y, Dicksved J, Riccardi G, Landberg R. The human gut microbiota and glucose metabolism: a scoping review of key bacteria and the potential role of SCFAs. *Am J Clin Nutr*. 2022 Oct 6;116(4):862-874.
- [145] Glucose alters the symbiotic relationships between gut microbiota and host physiology. Fernando F. Anhê, Nicole G. Barra, and Jonathan D. Schertzer *American Journal of Physiology-Endocrinology and Metabolism* 2020;318:2.E111-E116.
- [146] Dulaj, A.S.; Min, M.; Svamani, R.K. The Gut Microbiome's Influence on Incretins and Impact on Blood Glucose Control. *Biomedicines* 2024, 12, 2719.
- [147] Kristina M. Utzschneider, Mario Kratz, Chris J. Damman, Meredith Hullarg, Mechanisms Linking the Gut Microbiome and Glucose Metabolism, *The Journal of Clinical Endocrinology & Metabolism*, Volume 101, Issue 4, 1 April 2016, Pages 1445–1454
- [148] Poll BG, Cheema MU, Pluznick JL. Gut Microbial Metabolites and Blood Pressure Regulation: Focus on SCFAs and TMAO. *Physiology (Bethesda)*. 2020 Jul 1;35(4):275-284.
- [149] O'Donnell JA, Zheng T, Meric G, Marques FZ. The gut microbiome and hypertension. *Nat Rev Nephrol*. 2023 Mar;19(3):153-167.
- [150] Yang, Z., Wang, Q., Liu, Y., Wang, L., Ge, Z., Li, Z., ... Wu, C. (2023). Gut microbiota and hypertension: association, mechanisms and treatment. *Clinical and Experimental Hypertension*, 45(1).
- [151] Al Khodor S, Reichert B and Shatat IF (2017) The Microbiome and Blood Pressure: Can Microbes Regulate Our Blood Pressure? *Front. Pediatr*. 5:138.
- [152] Dinakis E, Nakai M, Gill P, Ribeiro R, Yiallourou S, Sata Y, Muir J, Carrington M, Head GA, Kaye DM, Marques FZ. Association Between the Gut Microbiome and Their Metabolites With Human Blood Pressure Variability. *Hypertension*. 2022 Aug;79(8):1690-1701.
- [153] Ge, Y., Wang, J., Wu, L., & Wu, J. (2024). Gut microbiota: A potential new regulator of hypertension. *Frontiers in Cardiovascular Medicine*, 11. *Frontiers | Gut microbiota: a potential new regulator of hypertension*
- [154] Hsu, C.L., Schnabl, B. The gut–liver axis and gut microbiota in health and liver disease. *Nat Rev Microbiol* 21, 719–733 (2023).
- [155] Tilg H, Adolph TE, Trauner M. Gut-liver axis: Pathophysiological concepts and clinical implications. *Cell Metab*. 2022 Nov 1;34(11):1700-1718.
- [156] Tripathi, A., Debellus, J., Brenner, D.A. et al. The gut–liver axis and the intersection with the microbiome. *Nat Rev Gastroenterol Hepatol* 15, 397–411 (2018).
- [157] Milosevic, I.; Vujovic, A.; Barac, A.; Djelic, M.; Korac, M.; Radovanovic Spunic, A.; Gmizic, I.; Stevanovic, O.; Djordjevic, V.; Lekic, N.; et al. Gut-Liver Axis, Gut Microbiota, and Its Modulation in the Management of Liver Diseases: A Review of the Literature. *Int. J. Mol. Sci*. 2019, 20, 395.
- [158] Fang L and Ning J (2024) Recent advances in gut microbiota and thyroid disease: pathogenesis and therapeutics in autoimmune, neoplastic, and nodular conditions. *Front. Cell. Infect. Microbiol*. 14:1465928.
- [159] Knezevic J, Starchi C, Tmava Berisha A, Amrein K. Thyroid-Gut-Axis: How Does the Microbiota Influence Thyroid Function? *Nutrients*. 2020 Jun 12;12(6):1769.
- [160] Xie L, Zhao H and Chen W (2023) Relationship between gut microbiota and thyroid function: a two-sample Mendelian randomization study. *Front. Endocrinol*. 14:1240752.
- [161] Sessa L, Malavolta E, Sodero G, Cipolla C, Rigante D. The conspiring role of gut microbiota as primer of autoimmune thyroid diseases: A scoping focus. *Autoimmun Rev*. 2025 Apr 30;24(5):103780.
- [162] Shi C, Chen J, He S, Zhang Y, Zhang Y and Yu L (2024) Cross-talk between the gut microbiota and hypothyroidism: a bidirectional two-sample Mendelian randomization study. *Front. Nutr*. 11:1286593. doi: 10.3389/fnut.2024.1286593

- [163] Wong, M., & Inder, W. J. (2018). Alternating hyperthyroidism and hypothyroidism in Graves' disease. Clinical case reports, 6(9), 1684–1688.
- [164] Mariat D, et al. The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. BMC Microbiol. 2009.
- [165] Magne F, et al. The Firmicutes/Bacteroidetes ratio: A relevant marker of gut dysbiosis in obese patients? Nutrients. 2020.
- [166] Stojanov S, et al. The influence of probiotics on the Firmicutes/Bacteroidetes ratio in the treatment of obesity and inflammatory bowel disease. Microorganisms. 2020.
- [167] Grigor'eva IN. Gallstone disease, obesity and the Firmicutes/Bacteroidetes ratio as a possible biomarker of gut dysbiosis. J Pers Med. 2021.
- [168] Holzapfel WH, et al. Taxonomy and important features of probiotic microorganisms in food and nutrition. Am J Clin Nutr. 2001.
- [169] Masco L, et al. Polyphasic taxonomic analysis of *Bifidobacterium animalis* and *Bifidobacterium lactis* reveals relatedness at the subspecies level: Reclassification of *Bifidobacterium animalis* as *Bifidobacterium animalis* subsp. *animalis* subsp. nov. and *Bifidobacterium lactis* as *Bifidobacterium animalis* subsp. *lactis* subsp. nov. Int J Syst Evol Microbiol. 2004.
- [170] Malinen E, et al. Association of symptoms with gastrointestinal microbiota in irritable bowel syndrome. World J Gastroenterol. 2010.
- [171] O'Callaghan A, et al. Bifidobacteria and their role as members of the human gut microbiota. Front Microbiol. 2016.
- [172] Rivière A, et al. Bifidobacteria and butyrate-producing colon bacteria: Importance and strategies for their stimulation in the human gut. Front Microbiol. 2016.
- [173] Marco ML, et al. Health benefits of fermented foods: Microbiota and beyond. Curr Opin Biotechnol. 2017.
- [174] Liu YW, et al. New perspectives of *Lactobacillus plantarum* as a probiotic: The gut-heart-brain axis. J Microbiol. 2018.
- [175] Heeney DD, et al. Intestinal *Lactobacillus* in health and disease: A driver or just along for the ride? Curr Opin Biotechnol. 2018.
- [176] Mayengbam S, et al. Impact of dietary fiber supplementation on modulating microbiota-host-metabolic axes in obesity. J Nutr Biochem. 2018.
- [177] Westerik N, et al. *Lactobacillus rhamnosus* probiotic food as a tool for empowerment across the value chain in Africa. Front Microbiol. 2018.
- [178] Capurso L, et al. Thirty years of *Lactobacillus rhamnosus* GG: A review. J Clin Gastroenterol. 2019.
- [179] Turroni F, et al. *Bifidobacterium bifidum*: A key member of the early human gut microbiota. Microorganisms. 2019.
- [180] Zhang T, et al. *Akkermansia muciniphila* is a promising probiotic. Microb Biotechnol. 2019.
- [181] Markowiak-Kopeć L, et al. The effect of probiotics on the production of short-chain fatty acids by the human intestinal microbiome. Nutrients. 2020.
- [182] Namova N, et al. Human gut microbiome response to short-term *Bifidobacterium*-based probiotic treatment. Indian J Microbiol. 2020.
- [183] Teame T, et al. Paraprobiotics and postbiotics of probiotic *Lactobacilli*, their positive effects on the host and action mechanisms: A review. Front Nutr. 2020.
- [184] Wang H, et al. The potential therapeutic role of *Lactobacillus reuteri* for treatment of inflammatory bowel disease. Am J Transl Res. 2020.
- [185] Bengoa AA, et al. Health-promoting properties of *Lactocaseibacillus paracasei*: A focus on kefir isolates and exopolysaccharide-producing strains. Foods. 2021.
- [186] Chen J, et al. Recent development of probiotic *Bifidobacteria* for treating human diseases. Front Bioeng Biotechnol. 2021.
- [187] Paul AK, et al. Probiotics and amelioration of rheumatoid arthritis: Significant roles of *Lactobacillus casei* and *Lactobacillus acidophilus*. Microorganisms. 2021.
- [188] Dempsey E, et al. *Lactobacillus* spp. for gastrointestinal health: Current and future perspectives. Front Immunol. 2022.
- [189] Gao H, et al. The functional roles of *Lactobacillus acidophilus* in different physiological and pathological processes. J Microbiol Biotechnol. 2022.
- [190] Luo Y, et al. Rational consideration of *Akkermansia muciniphila* targeting intestinal health: Advantages and challenges. npj Biofilms Microbiomes. 2022.
- [191] Echegaray N, et al. A novel approach to *Lactiplantibacillus plantarum*: From probiotic properties to the omics insights. Microbiol Res. 2023.
- [192] Guo H, et al. The potential therapeutic role of *Lactobacillaceae rhamnosus* for treatment of inflammatory bowel disease. Foods. 2023.
- [193] Yu Z, et al. The role of potential probiotic strains *Lactobacillus reuteri* in various intestinal diseases: New roles for an old player. Front Microbiol. 2023.
- [194] Huang JY, Lee SM, Mazmanian SK. The human commensal *Bacteroides fragilis* binds intestinal mucin. Anaerobe. 2011.
- [195] McDermott AJ, Huffnagle GB. The microbiome and regulation of mucosal immunity. Immunology. 2014.
- [196] Cornick S, Tawiah A, Chadee K. Roles and regulation of the mucus barrier in the gut. Tissue Barriers. 2015.
- [197] King SJ, McCole DF. Epithelial-microbial diplomacy: escalating border tensions drive inflammation in inflammatory bowel disease. Intest Res. 2019.
- [198] Qi H, et al. *Lactobacillus* maintains healthy gut mucosa by producing L-Ornithine. Commun Biol. 2019.
- [199] Paone P, Cani PD. Mucus barrier, mucins and gut microbiota: the expected slimy partners? Gut. 2020.
- [200] Fang J, et al. Slimy partners: the mucus barrier and gut microbiome in ulcerative colitis. Exp Mol Med. 2021.
- [201] Glover JS, et al. Characterizing the mucin-degrading capacity of the human gut microbiota. Sci Rep. 2022.
- [202] Lee JH, et al. A mucin-responsive hybrid two-component system controls *Bacteroides thetaiotaomicron* colonization and gut homeostasis. J Microbiol. 2022.
- [203] Liu MJ, et al. Recent findings in *Akkermansia muciniphila*-regulated metabolism and its role in intestinal diseases. Clin Nutr. 2022.
- [204] Rodrigues VF, et al. *Akkermansia muciniphila* and gut immune system: A good friendship that attenuates inflammatory bowel disease, obesity, and diabetes. Front Immunol. 2022.
- [205] Qin D, et al. Contribution of *Lactobacilli* on intestinal mucosal barrier and diseases: Perspectives and challenges of *Lactobacillus casei*. Life (Basel). 2022.
- [206] Gutierrez A, et al. *Bifidobacterium* and the intestinal mucus layer. Microbiome Res Rep. 2023.
- [207] He Q, et al. Protective effects of a new generation of probiotic *Bacteroides fragilis* against colitis in vivo and in vitro. Sci Rep. 2023.
- [208] Luis AS, Hansson GC. Intestinal mucus and their glycans: A habitat for thriving microbiota. Cell Host Microbe. 2023.
- [209] Pellegrino A, et al. Role of *Akkermansia* in human diseases: From causation to therapeutic properties. Nutrients. 2023.
- [210] Barcenilla A, et al. Phylogenetic relationships of butyrate-producing bacteria from the human gut. Appl Environ Microbiol. 2000.
- [211] Duncan SH, et al. Growth requirements and fermentation products of *Fusobacterium prausnitzii*, and a proposal to reclassify it as *Faecalibacterium prausnitzii* gen. nov., comb. nov. Int J Syst Evol Microbiol. 2002.
- [212] Schwierz A, et al. *Anaerostipes caecae* gen. nov., sp. nov., a new saccharolytic, acetate-utilising, butyrate-producing bacterium from human faeces. Syst Appl Microbiol. 2002.
- [213] Hold GL, et al. Oligonucleotide probes that detect quantitatively significant groups of butyrate-producing bacteria in human feces. Appl Environ Microbiol. 2003.
- [214] Duncan SH, et al. Contribution of acetate to butyrate formation by human faecal bacteria. Br J Nutr. 2004.
- [215] Duncan SH, et al. Lactate-utilizing bacteria, isolated from human feces, that produce butyrate as a major fermentation product. Appl Environ Microbiol. 2004.
- [216] Duncan SH, et al. Proposal of *Roseburia faecis* sp. nov., *Roseburia hominis* sp. nov. and *Roseburia inulinivorans* sp. nov., based on isolates from human faeces. Int J Syst Evol Microbiol. 2006.
- [217] Louis P, et al. Diversity, metabolism and microbial ecology of butyrate-producing bacteria from the human large intestine. FEMS Microbiol Lett. 2009.
- [218] Miquel S, et al. Ecology and metabolism of the beneficial intestinal commensal bacterium *Faecalibacterium prausnitzii*. Gut Microbes. 2014.
- [219] Miquel S, et al. Identification of metabolic signatures linked to anti-inflammatory effects of *Faecalibacterium prausnitzii*. MBio. 2015.
- [220] Engels C, et al. The common gut microbe *Eubacterium hallii* also contributes to intestinal propionate formation. Front Microbiol. 2016.
- [221] Rivière A, et al. Bifidobacteria and butyrate-producing colon bacteria: importance and strategies for their stimulation in the human gut. Front Microbiol. 2016.
- [222] Lopez-Siles M, et al. *Faecalibacterium prausnitzii*: from microbiology to diagnostics and prognostics. ISME J. 2017.
- [223] Tamani-Shacoori Z, et al. *Roseburia* spp.: A marker of health? Future Microbiol. 2017.
- [224] Bui TPN, et al. Mutual metabolic interactions in co-cultures of the intestinal *Anaerostipes rhamnosivorans* with an acetogen, methanogen, or pectin-degrader affecting butyrate production. Front Microbiol. 2019.
- [225] Chia LW, et al. *Bacteroides thetaiotaomicron* fosters the growth of butyrate-producing *Anaerostipes caecae* in the presence of lactose and total human milk carbohydrates. Microorganisms. 2020.
- [226] Van Hul M, et al. From correlation to causality: the case of *Subdoligranulum*. Gut Microbes. 2020.
- [227] Nie K, et al. *Roseburia intestinalis*: A beneficial gut organism from the discoveries in genus and species. Front Cell Infect Microbiol. 2021.
- [228] Singh V, et al. Butyrate producers, "The Sentinel of Gut": Their intestinal significance with and beyond butyrate, and prospective use as microbial therapeutics. Front Microbiol. 2023.
- [229] Notting F, et al. The butyrate-producing and spore-forming bacterial genus *Coprococcus* as a potential biomarker for neurological disorders. Gut Microbiome. 2023.
- [230] Kadowaki R, et al. Spore-forming properties and enhanced oxygen tolerance of butyrate-producing *Anaerostipes* spp. Anaerobe. 2023.
- [231] Siptroth J, et al. Variation of butyrate production in the gut microbiome in type 2 diabetes patients. Int Microbiol. 2023.
- [232] Gibson G, et al. Sulphate-reducing bacteria and hydrogen metabolism in the human large intestine. Gut. 1993.
- [233] Loubinoux J, et al. Sulfate-reducing bacteria in human feces and their association with inflammatory bowel diseases. FEMS Microbiol Ecol. 2002.
- [234] Jia W, et al. Diversity and distribution of sulphate-reducing bacteria in human faeces from healthy subjects and patients with inflammatory bowel disease. FEMS Immunol Med Microbiol. 2012.
- [235] Rey FE, et al. Metabolic niche of a prominent sulfate-reducing human gut bacterium. Proc Natl Acad Sci U S A. 2013.
- [236] David LA, et al. Diet rapidly and reproducibly alters the human gut microbiome. Nature. 2014.
- [237] Feng Y, et al. Enrichment of sulfidogenic bacteria from the human intestinal tract. FEMS Microbiol Lett. 2017.
- [238] Chen YR, et al. Isolation of *Desulfovibrio* spp. from human gut microbiota using a next-generation sequencing directed culture method. Lett Appl Microbiol. 2019.
- [239] Dostal Webster A, et al. Influence of short-term changes in dietary sulfur on the relative abundances of intestinal sulfate-reducing bacteria. Gut Microbes. 2019.
- [240] Kushkevych I, et al. The sulfate-reducing microbial communities and meta-analysis of their occurrence during diseases of small-large intestine axis. J Clin Med. 2019.
- [241] Dordovic D, et al. Hydrogen sulfide toxicity in the gut environment: Meta-analysis of sulfate-reducing and lactic acid bacteria in inflammatory processes. J Adv Res. 2020.
- [242] Kushkevych I, et al. Recent advances in metabolic pathways of sulfate reduction in intestinal bacteria. Cells. 2020.
- [243] Burrichter AG, et al. Bacterial microcompartments for isethionine desulfonation in the taurine-degrading human-gut bacterium *Bilophila wadsworthia*. BMC Microbiol. 2021.
- [244] Chen YR, et al. *Desulfovibrio* is not always associated with adverse health effects in the Guangdong Gut Microbiome Project. PeerJ. 2021.
- [245] Lu G, et al. Diversity and comparison of intestinal *Desulfovibrio* in patients with liver cirrhosis and healthy people. Microorganisms. 2023.
- [246] Singh SB, et al. *Desulfovibrio* in the gut: The enemy within? Microorganisms. 2023.
- [247] Xie R, et al. *Desulfovibrio vulgaris* interacts with novel gut epithelial immune receptor LRRC19 and exacerbates colitis. Microbiome. 2024.
- [248] Brown C, et al. The monoamine oxidase inhibitor—tyramine interaction. The Journal of Clinical Pharmacology. 1989.
- [249] Cummings JH, Macfarlane GT. Dietary fiber and the human gut microbiota. Nutrients. 1997.
- [250] Reiter RJ, Tan D. Melatonin: an antioxidant in edible plants. Annals of the New York Academy of Sciences. 2002.
- [251] Cherbut C, et al. *Acacia gum* is a bifidogenic dietary fibre with high digestive tolerance in healthy humans. Microbial Ecology in Health and Disease. 2003.
- [252] Iriti M, Faoro F. Grape phytochemicals: A bouquet of old and new nutraceuticals for human health. Medical Hypotheses. 2006.
- [253] Kinoshita T, et al. Cold-water face immersion per se elicits cardiac parasympathetic activity. Circulation Journal. 2006.

- [254] Reiter M, et al. Melatonin in edible plants (phytomelatonin): identification, concentrations, bioavailability and proposed functions. *World Review of Nutrition and Dietetics*. 2006.
- [255] Osman N, et al. Probiotics and blueberry attenuate the severity of dextran sulfate sodium (DSS)-induced colitis. *Digestive Diseases and Sciences*. 2008.
- [256] Holma R, et al. Constipation is relieved more by rye bread than wheat bread or laxatives without increased adverse gastrointestinal effects. *The Journal of Nutrition*. 2010.
- [257] Pérez-Jiménez J, et al. Identification of the 100 richest dietary sources of polyphenols: an application of the Phenol-Explorer database. *European Journal of Clinical Nutrition*. 2010.
- [258] Block SG, et al. Exacerbation of facial acne vulgaris after consuming pure chocolate. *Journal of the American Academy of Dermatology*. 2011.
- [259] Farinatti PT, et al. Acute effects of stretching exercise on the heart rate variability in subjects with low flexibility levels. *Journal of Strength and Conditioning Research*. 2011.
- [260] Russell WR, et al. High-protein, reduced-carbohydrate weight-loss diets promote metabolite profiles likely to be detrimental to colonic health. *The American Journal of Clinical Nutrition*. 2011.
- [261] Clemente JC, et al. The impact of the gut microbiota on human health: an integrative view. *Frontiers in Microbiology*. 2012.
- [262] Di Landro A, et al. Family history, body mass index, selected dietary factors, menstrual history, and risk of moderate to severe acne in adolescents and young adults. *Journal of the American Academy of Dermatology*. 2012.
- [263] Kim KA, et al. High-fat diet-induced gut microbiota exacerbates inflammation and obesity in mice via the TLR4 signaling pathway. *PLoS ONE*. 2012.
- [264] Queipo-Ortuño MI, et al. Influence of red wine polyphenols and ethanol on the gut microbiota ecology and biochemical biomarkers. *The American Journal of Clinical Nutrition*. 2012.
- [265] Biedermann L, et al. Bilberry ingestion improves disease activity in mild to moderate ulcerative colitis—an open pilot study. *Journal of Crohn's and Colitis*. 2013.
- [266] Delgado J, et al. Diets enriched with a Jerte Valley cherry-based nutraceutical product reinforce nocturnal behaviour in young and old animals of nocturnal (*Rattus norvegicus*) and diurnal (*Streptopelia risoria*) chronotypes. *Journal of Animal Physiology and Animal Nutrition*. 2013.
- [267] Dreher ML, Davenport AJ. Horticultural foods and health: the role of phytochemicals in nutrition and health promotion. *Journal of Horticultural Science & Biotechnology*. 2013.
- [268] Fritz JV, et al. The role of gut microbiota in metabolic syndrome: insights from genetic studies. *Nature Reviews Gastroenterology & Hepatology*. 2013.
- [269] Jelinek GA, et al. Association of fish consumption and omega-3 supplementation with quality of life, disability, and disease activity in an international cohort of people with multiple sclerosis. *International Journal of Neuroscience*. 2013.
- [270] Sae-Teaw M, et al. Serum melatonin levels and antioxidant capacities after consumption of pineapple, orange, or banana by healthy male volunteers. *Journal of Pineal Research*. 2013.
- [271] Spaeth AM, et al. Effects of experimental sleep restriction on weight gain, caloric intake, and meal timing in healthy adults. *Sleep*. 2013.
- [272] Festi D, et al. Gut microbiota and metabolic syndrome. *World Journal of Gastroenterology*. 2014.
- [273] Hansen AL, et al. Fish consumption, sleep, daily functioning, and heart rate variability. *Journal of Clinical Sleep Medicine*. 2014.
- [274] Jang SE, et al. Doenjang, a fermented Korean soybean paste, inhibits lipopolysaccharide production of gut microbiota in mice. *Journal of Medicinal Food*. 2014.
- [275] Katagiri R, et al. Low intake of vegetables, high intake of confectionary, and unhealthy eating habits are associated with poor sleep quality among middle-aged female Japanese workers. *Journal of Occupational Health*. 2014.
- [276] Panda S, et al. Structure and functions of the gut microbiome. *Endocrine, Metabolic & Immune Disorders-Drug Targets*. 2014.
- [277] Schoenaker DA, et al. The association between dietary factors and gestational hypertension and pre-eclampsia: a systematic review and meta-analysis of observational studies. *BMC Medicine*. 2014.
- [278] Chambers ES, et al. Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. *Gut*. 2015.
- [279] Guilloty NI, et al. Diet, pre-pregnancy BMI, and gestational weight gain in Puerto Rican women. *Maternal and Child Health Journal*. 2015.
- [280] Kavuri V, et al. Irritable bowel syndrome: Yoga as remedial therapy. *Evidence-Based Complementary and Alternative Medicine*. 2015.
- [281] Merckx A, et al. Weight gain in healthy pregnant women in relation to pre-pregnancy BMI, diet and physical activity. *Midwifery*. 2015.
- [282] Pagliaro B, et al. Phytochemical compounds and protection from cardiovascular diseases: a state of the art. *BioMed Research International*. 2015.
- [283] Schmidt K, et al. Probiotic intake reduces the waking cortisol response and alters emotional bias in healthy volunteers. *Psychopharmacology*. 2015.
- [284] Singh CK, et al. Resveratrol and cancer: challenges for clinical translation. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*. 2015.
- [285] Shin D, et al. Dietary patterns during pregnancy are associated with risk of gestational diabetes mellitus. *Nutrients*. 2015.
- [286] Zulueta A, et al. Resveratrol: A potential challenger against gastric cancer. *World Journal of Gastroenterology*. 2015.
- [287] Braune A, et al. Bacterial species involved in the conversion of dietary flavonoids in the human gut. *Gut Microbes*. 2016.
- [288] Chausse B, et al. Microbiota modulation by dietary fibers: what is known and what needs to be elucidated? *Nutrients*. 2016.
- [289] Delost GR, et al. The impact of chocolate consumption on acne vulgaris in college students: A randomized crossover study. *Journal of the American Academy of Dermatology*. 2016.
- [290] Di Landro A, et al. Adult female acne and associated risk factors: Results of a multicenter case-control study in Italy. *Journal of the American Academy of Dermatology*. 2016.
- [291] Grossi E, et al. The constellation of dietary factors in adolescent acne: A semantic connectivity map approach. *Journal of the European Academy of Dermatology and Venereology*. 2016.
- [292] Konstantinidou, et al. Cardiorespiratory responses and reduced apneic time to cold-water face immersion after high intensity exercise. *Respiratory Physiology & Neurobiology*. 2016.
- [293] Kuršvietienė L, et al. Multiplicity of effects and health benefits of resveratrol. *Medicina*. 2016.
- [294] Roth S, et al. Bilberry-derived anthocyanins modulate cytokine expression in the intestine of patients with ulcerative colitis. *PLOS ONE*. 2016.
- [295] Turan N and Atabek Aş T. The effect of abdominal massage on constipation and quality of life. *Gastroenterology Nursing*. 2016.
- [296] Vongraviopap S and Asawanonda P. Dark chocolate exacerbates acne. *International Journal of Dermatology*. 2016.
- [297] Ziegler JU, et al. Wheat and the irritable bowel syndrome – FODMAP levels of modern and ancient species and their retention during bread making. *Journal of Functional Foods*. 2016.
- [298] Chekroud AM, et al. Relationship between dietary quality and gut microbiota in humans. *Nutrients*. 2017.
- [299] Fuller C, et al. Bedtime use of technology and associated sleep problems in children. *Global Pediatric Health*. 2017.
- [300] Gomes da Silva MF et al. Yacon syrup: food applications and impact on satiety in healthy volunteers. *Food Research International*. 2017.
- [301] Garcia-Mazcorro JF, et al. Gut microbiome modulation by fermented foods. *Frontiers in Microbiology*. 2017.
- [302] Gibson GR, et al. Dietary prebiotics: definition and application. *Nature Reviews Gastroenterology & Hepatology*. 2017.
- [303] Hewlings S, Kalman D. Curcumin: A review of its effects on human health. *Foods*. 2017.
- [304] Johnson EL, et al. Microbiome and metabolic disease: revisiting the bacterial phylum Bacteroidetes. *Journal of Molecular Medicine*. 2017.
- [305] Maier TV, et al. Impact of dietary resistant starch on the human gut microbiome, metaproteome, and metabolome. *mBio*. 2017.
- [306] Reynolds AC, et al. The shift work and health research agenda: considering changes in gut microbiota as a pathway linking shift work, sleep loss and circadian misalignment, and metabolic disease. *Sleep Medicine Reviews*. 2017.
- [307] Vandeputte D, et al. Probiotic inulin-type fructans induce specific changes in the human gut microbiota. *Gut*. 2017.
- [308] Yuen AWC and Sander JW. Can natural ways to stimulate the vagus nerve improve seizure control? *Epilepsy & Behavior*. 2017.
- [309] Adike A, et al. Small intestinal bacterial overgrowth. *Gastroenterology Clinics of North America*. 2018.
- [310] Alfa MJ, et al. A randomized trial to determine the impact of a digestion resistant starch composition on the gut microbiome in older and mid-age adults. *Clinical Nutrition*. 2018.
- [311] Durack J, et al. Involvement of the gut microbiome in diabetes and obesity. *International Journal of Molecular Sciences*. 2018.
- [312] Gao Q, et al. The association between vitamin D deficiency and sleep disorders: A systematic review and meta-analysis. *Nutrients*. 2018.
- [313] Ghosh S, Maji S. Dietary approaches to improve gut health. *Journal of Nutritional Science*. 2018.
- [314] González-Sarrías A, et al. The endotoxemia marker lipopolysaccharide-binding protein is reduced in overweight-obese subjects consuming pomegranate extract by modulating the gut microbiota: A randomized clinical trial. *Molecular Nutrition & Food Research*. 2018.
- [315] Hershner S, et al. The impact of a randomized sleep education intervention for college students. *Journal of Clinical Sleep Medicine*. 2018.
- [316] Kopf JC, et al. Role of whole grains versus fruits and vegetables in reducing subclinical inflammation and promoting gastrointestinal health in individuals affected by overweight and obesity: a randomized controlled trial. *Nutrition Journal*. 2018.
- [317] Korpela K, et al. Diet, microbiota, and metabolic health: trade-off between saccharolytic and proteolytic fermentation. *Annual Review of Food Science and Technology*. 2018.
- [318] Krebs-Smith SM, et al. Update of the Healthy Eating Index: HEI-2015. *Journal of the Academy of Nutrition and Dietetics*. 2018.
- [319] Niederreiter L, et al. Food, microbiome and colorectal cancer. *Digestive and Liver Disease*. 2018.
- [320] Schink M, et al. Microbial patterns in patients with histamine intolerance. *Journal of Physiology and Pharmacology*. 2018.
- [321] Venancio VP, et al. Polyphenol-rich mango (*Mangifera indica* L.) ameliorates functional constipation symptoms in humans beyond equivalent amount of fiber. *Molecular Nutrition & Food Research*. 2018.
- [322] Vital M, et al. Metagenomic insights into the degradation of resistant starch by human gut microbiota. *Applied and Environmental Microbiology*. 2018.
- [323] Aalemi AK, et al. Dairy consumption and acne: a case control study in Kabul, Afghanistan. *Clinical, Cosmetic and Investigational Dermatology*. 2019.
- [324] Chaix A, et al. Time-restricted eating to prevent and manage chronic metabolic diseases. *Annual Review of Nutrition*. 2019.
- [325] Chung WH, et al. Dietary factors influencing the gut microbiome in children: a systematic review. *Nutrients*. 2019.
- [326] Clos-García M, et al. Gut microbiome and serum metabolome analyses identify molecular biomarkers and altered glutamate metabolism in fibromyalgia. *eBioMedicine*. 2019.
- [327] Covasa M, et al. Intestinal sensing by gut microbiota: Targeting gut peptides. *Frontiers in Endocrinology*. 2019.
- [328] Dalton A, et al. Exercise influence on the microbiome—gut—brain axis. *Gut Microbes*. 2019.
- [329] Danneskiold-Samsøe NB, et al. Interplay between food and gut microbiota in health and disease. *Food Research International*. 2019.
- [330] Erdogan M, et al. The impact of dietary fat on gut microbiome: a systematic review. *Journal of Nutrition and Metabolism*. 2019.
- [331] Hermann J. Drug-nutrient interactions. 2019.
- [332] Hills R, et al. Gut microbiome: Profound implications for diet and disease. *Nutrients*. 2019.
- [333] Hjørth MF, et al. Prevotella-to-Bacteroides ratio predicts body weight and fat loss success on 24-week diets varying in macronutrient composition and dietary fiber: results from a post-hoc analysis. *International Journal of Obesity*. 2019.
- [334] Hrnčirova L, et al. Human gut microbes are susceptible to antimicrobial food additives in vitro. *Folia Microbiologica*. 2019.
- [335] Kaczmarek JL, et al. Broccoli consumption affects the human gastrointestinal microbiota. *The Journal of Nutritional Biochemistry*. 2019.
- [336] Kanauchi M, et al. A novel dietary inflammatory index reflecting inflammatory ageing: Technical note. *Annals of Medicine and Surgery*. 2019.
- [337] Kårlund A, et al. Protein supplements and their relation with nutrition, microbiota composition, and health: is more protein always better for sportspeople? *Nutrients*. 2019.

- [338] Partula V, et al. Associations between usual diet and gut microbiota composition: results from the Milieu Intérieur cross-sectional study. *The American Journal of Clinical Nutrition*. 2019.
- [339] Rao TP, et al. Role of guar fiber in improving digestive health and function. *Nutrition*. 2019.
- [340] Saeji AA, et al. Effects of yogic breath regulation: A narrative review of scientific evidence. *Journal of Ayurveda and Integrative Medicine*. 2019.
- [341] Zhao J, et al. Dietary protein and gut microbiota composition and function. *Current Protein & Peptide Science*. 2019.
- [342] Ahnen RT, et al. Carbohydrates. *Present Knowledge in Nutrition*. 2020.
- [343] Atzeni A, et al. Exercise, diet and stress as modulators of gut microbiota: implications for neurodegenerative diseases. *Neurobiology of Disease*. 2020.
- [344] Ahnen RT, et al. Carbohydrates. In: *Present Knowledge in Nutrition*. Elsevier. 2020.
- [345] Bonci L, et al. Diet strategies for managing chronic diarrhea. *International Foundation for Gastrointestinal Disorders*. 2020.
- [346] Cándido TLN, et al. Effects of dietary fat quality on metabolic endotoxaemia: a systematic review. *British Journal of Nutrition*. 2020.
- [347] Chou SW, et al. Dietary fibers in the treatment of constipation: a review of randomized controlled trials. *Nutrients*. 2020.
- [348] Chung WSF, et al. Relative abundance of the *Prevotella* genus within the human gut microbiota of elderly volunteers determines the inter-individual responses to dietary supplementation with wheat bran arabinoxylan-oligosaccharides. *BMC Microbiology*. 2020.
- [349] Coombes JR, et al. Influence of dietary fat on gut microbiome composition and microbial metabolites. *Current Opinion in Clinical Nutrition and Metabolic Care*. 2020.
- [350] Davis CD, et al. Prebiotic dietary fiber modulates gut microbiota and metabolite profiles in prediabetic individuals. *Nutrients*. 2020.
- [351] Dionisio RP, et al. Effect of yacon syrup on blood lipid, glucose and metabolic endotoxaemia in healthy subjects: a randomized, double-blind, placebo-controlled pilot trial. *Food Science and Technology*. 2020.
- [352] Gubert C, et al. Exercise, diet and stress as modulators of gut microbiota: implications for neurodegenerative diseases. *Neurobiology of Disease*. 2020.
- [353] Gurwara S, et al. Alcohol use alters the colonic mucosa-associated gut microbiota in humans. *Nutrition Research*. 2020.
- [354] Hedström AK, et al. Low fish consumption is associated with a small increased risk of MS. *Neurology Neuroimmunology & Neuroinflammation*. 2020.
- [355] Hjorth MF, et al. Pretreatment *Prevotella*-to-*Bacteroides* ratio and salivary amylase gene copy number as prognostic markers for dietary weight loss. *The American Journal of Clinical Nutrition*. 2020.
- [356] Janczy A, et al. Impact of diet and synbiotics on selected gut bacteria and intestinal permeability in individuals with excess body weight – a prospective, randomized study. *Acta Biochimica Polonica*. 2020.
- [357] Labenz J, et al. Gastroösophageale Refluxkrankheit – update 2021. *Der Internist*. 2020.
- [358] Lee SH, et al. Emotional well-being and gut microbiome profiles by enterotype. *Scientific Reports*. 2020.
- [359] Machate DJ, et al. Fatty acid diets: Regulation of gut microbiota composition and obesity and its related metabolic dysbiosis. *International Journal of Molecular Sciences*. 2020.
- [360] Simiuc R, Turcanu D. Certain aspects of nutritional security of people with gluten-related disorders. *Food and Nutrition Sciences*. 2020.
- [361] Thomsen BJ, et al. The potential uses of omega-3 fatty acids in dermatology: a review. *Journal of Cutaneous Medicine and Surgery*. 2020.
- [362] Yee BE, et al. Serum zinc levels and efficacy of zinc treatment in acne vulgaris: a systematic review and meta-analysis. *Dermatologic Therapy*. 2020.
- [363] Zou H, et al. Effect of caloric restriction on BMI, gut microbiota, and blood amino acid levels in non-obese adults. *Nutrients*. 2020.
- [364] Die besten Hausmittel von A bis Z. Alternative Heilmittel aus der Hausapotheke der Natur, inklusive der Heilmethoden der Homöopathie, Akupunktur und des Tai Chi. Bassermann Verlag. 2020.
- [365] Beam A, et al. Effect of diet and dietary components on the composition of the gut microbiota. *Nutrients*. 2021.
- [366] Borsani B, et al. The role of carrageenan in inflammatory bowel diseases and allergic reactions: Where do we stand? *Nutrients*. 2021.
- [367] Clauss M, et al. Interplay between exercise and gut microbiome in the context of human health and performance. *Frontiers in Nutrition*. 2021.
- [368] Decker B, et al. Dietary fibers as prebiotics: mechanisms of action and health benefits. *Nutrients*. 2021.
- [369] Deshpande S, et al. Effect of dietary interventions on gut microbiota and their implications for health. *Molecules*. 2021.
- [370] Duffy M, et al. Gut microbiome, diet, and chronic disease. *Frontiers in Nutrition*. 2021.
- [371] Fachgesellschaft für Ernährungstherapie und Prävention (FET) eV. 2021.
- [372] Fu T, et al. Fermentation of alginate and its derivatives by different enterotypes of human gut microbiota: Towards personalized nutrition using enterotype-specific dietary fibers. *International Journal of Biological Macromolecules*. 2021.
- [373] Hahne D, et al. Dietary fiber and gut health. *Current Gastroenterology Reports*. 2021.
- [374] Hrubisko M, et al. Histamine intolerance—the more we know the less we know: A review. *Nutrients*. 2021.
- [375] Hosseini-Asl MK, et al. The effect of a short-term physical activity after meals on gastrointestinal symptoms in individuals with functional abdominal bloating: a randomized clinical trial. *Gastroenterology and Hepatology from Bed to Bench*. 2021.
- [376] Kolanos R, et al. German chamomile, Nutraceuticals. 2021.
- [377] Larson R, et al. Acacia gum is well tolerated while increasing satiety and lowering peak blood glucose response in healthy human subjects. *Nutrients*. 2021.
- [378] McQuilken SA, et al. Gut motility and its control. *Anaesthesia & Intensive Care Medicine*. 2021.
- [379] O'Brien L, et al. What are the pearls and pitfalls of the dietary management for chronic diarrhoea? *Nutrients*. 2021.
- [380] Pham VT, et al. Vitamins, the gut microbiome and gastrointestinal health in humans. *Nutrition Research*. 2021.
- [381] Podgórska A, et al. Acne vulgaris and intake of selected dietary nutrients—a summary of information. *Healthcare*. 2021.
- [382] Prasodanan PK, et al. Western and non-western gut microbiomes reveal new roles of *Prevotella* in carbohydrate metabolism and mouth–gut axis. *npj Biofilms and Microbiomes*. 2021.
- [383] Schnedl WJ, et al. Histamine intolerance originates in the gut. *Nutrients*. 2021.
- [384] Seo K, et al. The role of mucosal barriers in human gut health. *Archives of Pharmacol Research*. 2021.
- [385] Shil A, et al. Artificial sweeteners negatively regulate pathogenic characteristics of two model gut bacteria, *E. coli* and *E. faecalis*. *International Journal of Molecular Sciences*. 2021.
- [386] Srisukthaveerat V, et al. Nutrition communication about low FODMAP diet in irritable bowel syndrome (IBS) and small intestinal bacterial overgrowth (SIBO) in Thai healthcare practitioners. *Bioactive Compounds in Health and Disease*. 2021.
- [387] Sullivan VK, et al. Consumption of dried fruits is associated with greater intakes of underconsumed nutrients, higher total energy intakes, and better diet quality in US adults: a cross-sectional analysis of the National Health and Nutrition Examination Survey, 2007–2016. *Journal of the Academy of Nutrition and Dietetics*. 2021.
- [388] Wong A and Figueroa A. Effects of acute stretching exercise and training on heart rate variability: A review. *Journal of Strength and Conditioning Research*. 2021.
- [389] Watanabe-Asaka T, et al. From digestion and absorption to innate immunity and health care: Water and food intake may contribute to IL-22 in ILC3-dependent mucosal immunity in the jejunum. *The Journal of Physiological Sciences*. 2021.
- [390] Atzeni A, et al. Association between ultra-processed food consumption and gut microbiota in senior subjects with overweight/obesity and metabolic syndrome. *Frontiers in Nutrition*. 2022.
- [391] Bartlett A, et al. Dietary protein and the intestinal microbiota: an understudied relationship. *iScience*. 2022.
- [392] Cataldi S, et al. The relationship between physical activity, physical exercise, and human gut microbiota in healthy and unhealthy subjects: A systematic review. *Biology*. 2022.
- [393] Chassaing B, et al. Randomized controlled-feeding study of dietary emulsifier carboxymethylcellulose reveals detrimental impacts on the gut microbiota and metabolome. *Gastroenterology*. 2022.
- [394] Choi Y, et al. A guide to dietary pattern–microbiome data integration. *The Journal of Nutrition*. 2022.
- [395] Conforti C, et al. Acne and diet: a review. *International Journal of Dermatology*. 2022.
- [396] Davis KF, et al. Prebiotics: what they are, health benefits and food sources. *Molecules*. 2022.
- [397] Galle A, et al. Dietary interventions for the management of patients with inflammatory bowel diseases: a systematic review. *Nutrients*. 2022.
- [398] Gaundal L, et al. Gut microbiota is associated with dietary intake and metabolic markers in healthy individuals. *Food & Nutrition Research*. 2022.
- [399] González M, et al. Effects of probiotic and prebiotic interventions on gut microbiota and immune response: a systematic review. *Nutrients*. 2022.
- [400] Gupta CC, et al. A time to rest, a time to dine: Sleep, time-restricted eating, and cardiometabolic health. *Nutrients*. 2022.
- [401] Hur HJ, et al. Beneficial effects of a low-glycemic diet on serum metabolites and gut microbiota in obese women with *Prevotella* and *Bacteroides* enterotypes: a randomized clinical trial. *Frontiers in Nutrition*. 2022.
- [402] Iao SI, et al. Associations between bedtime eating or drinking, sleep duration and wake after sleep onset: findings from the American time use survey. *British Journal of Nutrition*. 2022.
- [403] Kyaw TS, et al. Monosodium glutamate consumption reduces the renal excretion of trimethylamine N-oxide and the abundance of *Akkermansia muciniphila* in the gut. *Biochemical and Biophysical Research Communications*. 2022.
- [404] Lakshmanan AP, et al. Increased relative abundance of *Ruminococcus* is associated with reduced cardiovascular risk in an obese population. *Frontiers in Nutrition*. 2022.
- [405] Marques C, et al. Impact of beer and nonalcoholic beer consumption on the gut microbiota: a randomized, double-blind, controlled trial. *Journal of Agricultural and Food Chemistry*. 2022.
- [406] Moniaga CS, Tominaga M, Takamori K. An altered skin and gut microbiota are involved in the modulation of itch in atopic dermatitis. *Cells*. 2022.
- [407] Pattnaik H, et al. Nutritional elements in sleep. *Currents*. 2022.
- [408] Phillips Michael M MD, et al. Abdominal bloating. *MedlinePlus Medical Encyclopedia*. 2022.
- [409] Sánchez-Pérez S, et al. Intestinal dysbiosis in patients with histamine intolerance. *Nutrients*. 2022.
- [410] Sanchez J, et al. Gastroesophageal reflux disease. *Interventional Management of Chronic Visceral Pain Syndromes*. 2022.
- [411] Yuan L, et al. Profile of hemoglobin, glucose, and triglycerides on the use of yacon prebiotic syrup as a natural supplement. *RASAYAN Journal of Chemistry*. 2022.
- [412] Yuan MZ, et al. Research on the impact of regular exercise behavior of college students on academic stress and sleep quality during the COVID-19 pandemic. *Healthcare (Basel)*. 2022.
- [413] Abe A, et al. Partially hydrolyzed guar gum is associated with improvement in gut health, sleep, and motivation among healthy subjects. *Journal of Clinical Biochemistry and Nutrition*. 2023.
- [414] Abdi F, et al. Nutritional considerations in celiac disease and non-celiac gluten/wheat sensitivity. *Nutrients*. 2023.
- [415] Alasalvar C, et al. Dried fruits: bioactives, effects on gut microbiota, and possible health benefits—an update. *Nutrients*. 2023.
- [416] Aleman RS, et al. Leaky gut and the ingredients that help treat it: a review. *Molecules*. 2023.
- [417] AOK. Was sollte man bei Durchfall (Diarrhoe) essen und trinken? AOK Gesundheitsmagazin. 2023.
- [418] Atzeni A, et al. Carbohydrate quality, fecal microbiota and cardiometabolic health in older adults: a cohort study. *Gut Microbes*. 2023.
- [419] Barber C, et al. Metabolic response of intestinal microbiota to guar gum consumption. *Frontiers in Nutrition*. 2023.
- [420] Bartsch M, et al. Bridging the gap from enterotypes to personalized dietary recommendations: a metabolomics perspective on microbiome research. *Metabolites*. 2023.
- [421] Benamer T, et al. The effects of curcumin on inflammasome: latest update. *Molecules*. 2023.
- [422] Bielik V, et al. The effect of physical exercise and dairy probiotics (*Lactobacillus casei*) on gut microbiome in childhood cancer survivors. *Neoplasma*. 2023.

- [423] Carvalho RDOD, et al. Functional Swiss-type cheeses promote beneficial effects in mice gut microbiome during homeostasis and inflammation. *Food Bioscience*. 2023.
- [424] Conesa MPB, et al. Stabilizing histamine release in gut mast cells mitigates peripheral and central inflammation after stroke. *Journal of Neuroinflammation*. 2023.
- [425] Dicks L MT, et al. Our mental health is determined by an intrinsic interplay between the central nervous system, enteric nerves, and gut microbiota. *International Journal of Molecular Sciences*. 2023.
- [426] Dempsey KW, et al. Wheat-derived fructans as prebiotics. *Food Research International*. 2023.
- [427] De Souza Lopes A, et al. The impact of antimicrobial food additives and sweeteners on the growth and metabolite production of gut bacteria. *Folia Microbiologica*. 2023.
- [428] Dicks LM, et al. Our mental health is determined by an intrinsic interplay between the central nervous system, enteric nerves, and gut microbiota. *International Journal of Molecular Sciences*. 2023.
- [429] Escobar LA, et al. The effect of soluble dietary fibers on gut microbiota: a systematic review and meta-analysis. *Frontiers in Nutrition*. 2023.
- [430] Fischer C, et al. Impact of dietary habits on gut microbiota and health: a systematic review. *Nutrients*. 2023.
- [431] Foshati S, et al. The effects of ginger supplementation on common gastrointestinal symptoms in patients with relapsing-remitting multiple sclerosis: A double-blind randomized placebo-controlled trial. *BMC Complementary Medicine and Therapies*. 2023.
- [432] Ghosh S, et al. Dietary polyphenols and their effects on gut microbiota in obesity: a review. *Food Chemistry*. 2023.
- [433] González-Morales J, et al. Prebiotic dietary fibers: a review of health benefits. *Nutrients*. 2023.
- [434] González-Orozco BD, et al. Metagenomic analysis and antibacterial activity of kefir microorganisms. *Journal of Food Science*. 2023.
- [435] González-Revelta I, et al. Fermented foods as potential modulators of the gut microbiota: a review. *Nutrients*. 2023.
- [436] Gostimirovic M, et al. Resveratrol and gut microbiota synergy: Preventive and therapeutic effects. *International Journal of Molecular Sciences*. 2023.
- [437] Griffiths A, et al. Effects of resistant starch on gut microbiota: a systematic review. *Nutrients*. 2023.
- [438] Gudan A, et al. Small intestinal bacterial overgrowth and non-alcoholic fatty liver disease: What do we know in 2023? *Nutrients*. 2023.
- [439] Hald BJ, et al. The role of the gut microbiome in obesity: recent advances. *Nature Reviews Endocrinology*. 2023.
- [440] Harvard MS, et al. What to eat when you have chronic heartburn. *Harvard Health Publishing*. 2023.
- [441] Herdiana Y, et al. Functional food in relation to gastroesophageal reflux disease (GERD). *Nutrients*. 2023.
- [442] Isaakidis A, et al. Is there more to olive oil than healthy lipids? *Nutrients*. 2023.
- [443] Kanwal F, et al. The potential role of nondigestible raffinose family oligosaccharides as prebiotics. *Glycobiology*. 2023.
- [444] Li A, et al. Multi-omics analyses reveal relationships among polyphenol-rich oolong tea consumption, gut microbiota, and metabolic profile: a pilot study. *Food Chemistry*. 2023.
- [445] Li XY, et al. Regulation of gut microbiota by vitamin C, vitamin E and β -carotene. *Food Research International*. 2023.
- [446] Mitra S, et al. Brain modulation by the gut microbiota: from disease to therapy. *Journal of Advanced Research*. 2023.
- [447] Ogulur I, et al. Mechanisms of gut epithelial barrier impairment caused by food emulsifiers polysorbate 20 and polysorbate 80. *Allergy*. 2023.
- [448] Pang S, et al. Longevity of centenarians is reflected by the gut microbiome with youth-associated signatures. *Nature Aging*. 2023.
- [449] Poto R, et al. The role of gut microbiota and leaky gut in the pathogenesis of food allergy. *Nutrients*. 2023.
- [450] Radziszewska M, et al. Nutrition, physical activity and supplementation in irritable bowel syndrome. *Nutrients*. 2023.
- [451] Sánchez-Terrón G, et al. Impact of sustained fructose consumption on gastrointestinal function and health in Wistar rats: glycoxidative stress, impaired protein digestion, and shifted fecal microbiota. *Journal of Agricultural and Food Chemistry*. 2023.
- [452] Scarpellini E, et al. The use of peppermint oil in gastroenterology. *Current Pharmaceutical Design*. 2023.
- [453] Schneidl WJ, et al. A personalized management approach in disorders of the irritable bowel syndrome spectrum. *Clinical Nutrition ESPEN*. 2023.
- [454] Sharifan P, et al. Association of dietary and blood inflammatory indicators with depression, anxiety, and stress in adults with vitamin D deficiency. *International Journal of Geriatric Psychiatry*. 2023.
- [455] Shevchenko A, et al. Post-stress changes in the gut microbiome composition in rats with different levels of nervous system excitability. *PLOS ONE*. 2023.
- [456] Simão DO, et al. Lipids, gut microbiota, and the complex relationship with Alzheimer's disease: a narrative review. *Nutrients*. 2023.
- [457] Stolz R, et al. Evidenzbasierte naturheilkundliche Pflegeinterventionen in der Schmerztherapie. *Der Schmerz*. 2023.
- [458] Trivedi G, et al. Humming (simple bhrumari pranayama) as a stress buster: A holter-based study to analyze heart rate variability (HRV) parameters during bhrumari, physical activity, emotional stress, and sleep. *Cureus*. 2023.
- [459] Wang, Y, et al. Effect of two-week red beetroot juice consumption on modulation of gut microbiota in healthy human volunteers – A pilot study. *Food Chemistry*. 2023.
- [460] Zhang X, et al. Modulating a prebiotic food source influences inflammation and immune-regulating gut microbes and metabolites: insights from the BE GONE trial. *eBioMedicine*. 2023.
- [461] Zhao Q, et al. The relationship between the dietary inflammatory index (DII) and metabolic syndrome (MetS) in middle-aged and elderly individuals in the United States. *Nutrients*. 2023.
- [462] Zhong HJ, et al. Supplementation with high-GABA-producing *Lactobacillus plantarum* L5 ameliorates essential tremor triggered by decreased gut bacteria-derived GABA. *Translational Neurodegeneration*. 2023.
- [463] Aguayo-Guerrero JA, et al. Sucralose: from sweet success to metabolic controversies—unraveling the global health implications of a pervasive non-caloric artificial sweetener. *Life*. 2024.
- [464] Ardisson Korat AV, et al. Dietary protein intake in midlife in relation to healthy aging – results from the prospective Nurses' Health Study cohort. *The American Journal of Clinical Nutrition*. 2024.
- [465] Bez NS, et al. Development of a diet quality score and adherence to the Swiss dietary recommendations for vegans. *Journal of Health, Population and Nutrition*. 2024.
- [466] Can G, et al. Unraveling the potential therapeutic effect of synbiotics in inflammatory bowel disease. *Journal of the Academy of Nutrition and Dietetics*. 2024.
- [467] Chatelan A, et al. Substituting low-calorie sweetened beverages for sugar-sweetened beverages to prevent obesity and cardiometabolic diseases: still a good idea? *Current Developments in Nutrition*. 2024.
- [468] Chen C, et al. *Lactobacillus paracasei* AH2 isolated from Chinese sourdough alleviated gluten-induced food allergy through modulating gut microbiota and promoting short-chain fatty acid accumulation in a BALB/c mouse model. *Journal of the Science of Food and Agriculture*. 2024.
- [469] Del Rosso J, et al. New insights into systemic drivers of inflammation and their contributions to the pathophysiology of acne. *Journal of Drugs in Dermatology*. 2024.
- [470] Hayer SS, et al. Antibiotic-induced gut dysbiosis elicits gut-brain axis relevant multi-omic signatures and behavioral and neuroendocrine changes in a nonhuman primate model. *Gut Microbes*. 2024.
- [471] Huang Y, et al. The adjuvant treatment role of ω -3 fatty acids by regulating gut microbiota positively in acne vulgaris. *Journal of Dermatological Treatment*. 2024.
- [472] Jamieson PE, et al. Gut enterotype-dependent modulation of gut microbiota and their metabolism in response to xanthohumol supplementation in healthy adults. *Gut Microbes*. 2024.
- [473] Konstanti P, et al. Physiology of γ -aminobutyric acid production by *Akkermansia muciniphila*. *Applied and Environmental Microbiology*. 2024.
- [474] Malta FAP, et al. A triple-masked, two-center, randomized parallel clinical trial to assess the superiority of eight weeks of grape seed flour supplementation against placebo for weight loss attenuation during perioperative period in patients with cachexia associated with colorectal cancer: a study protocol. *Frontiers in Endocrinology*. 2024.
- [475] Min L, et al. Effects of exercise on gut microbiota of adults: a systematic review and meta-analysis. *Nutrients*. 2024.
- [476] Mutoh N, et al. *Bifidobacterium breve* M-16V regulates the autonomic nervous system via the intestinal environment: A double-blind, placebo-controlled study. *Behavioural Brain Research*. 2024.
- [477] Palma-Ordóñez JF, et al. Implication of intestinal microbiota in the etiopathogenesis of fibromyalgia: a systematic review. *International Journal of Rheumatic Diseases*. 2024.
- [478] Pérez-Castillo IM, et al. The athlete gut microbiota: state of the art and practical guidance. *Beneficial Microbes*. 2024.
- [479] Pierdomenico M, et al. Anti-inflammatory effect of a pomegranate extract on LPS-stimulated HepG2 cells. *Natural Product Research*. 2024.
- [480] Prescott SL, et al. Nutritional criminology: Why the emerging research on ultra-processed food matters to health and justice. *International Journal of Environmental Research and Public Health*. 2024.
- [481] Weng J, et al. Gender differences in the association between healthy eating index-2015 and hypertension in the US population: evidence from NHANES 1999–2018. *BMC Public Health*. 2024.
- [482] WHO. Diarrhoea, WHO health topics. 2024.
- [483] Yadav SS, et al. Effect of yoga-based breathing practices on depression, anxiety, stress, and fear of COVID-19 positive hospitalized patients: A randomized controlled trial. *Journal of Ayurveda and Integrative Medicine*. 2024.